

Applying a Systematic Conservation-Planning Tool with Real Data of Canton Aargau



A part of the conservation area Auenschachen near Brugg AG; Photo: L.Götz

Master thesis

Luzia Götz

Master Environmental Science, Tödistrasse 17, 8620 Wetzikon, lgoetz@student.ethz.ch

Supervisors:

Dr. Peter B. Pearman
Land Use Dynamics, Swiss Federal Research Institute WSL, Birmensdorf

Prof. Dr. Felix Kienast
Landscape Dynamics, Swiss Federal Research Institute WSL, Birmensdorf

Birmensdorf, March 2014

Abstract

Systematic conservation planning is a contemporary issue. Since its origins in the early 1980s, a lot of research has been done in this field. One research sector has dealt with computerbased planning tools. From there, Marxan as a systematic conservation-planning tool has emerged. This tool operates as a decision support tool by finding optimal reserve systems given certain, defined constraints. However, while Marxan is possibly the most widely used systematic conservation-planning tool, it almost has rarely been applied in Switzerland. Hence, this master thesis was a first attempt to use a systematic conservation-planning tool with real plant and animal species data on a cantonal scale.

Several important conclusions arise from this study. First, Marxan's selection of conservation areas is reasonable and comprehensible. Second, having adequate input data is a critical issue. Third, applying Marxan as optimization software is only a part of the comprehensive concept of systematic conservation planning. Finally, this concept is holding a big potential for future planning processes in Switzerland.

Content

Abstract	III
1. Introduction.....	1
2. Methods.....	3
2.1 Study Site.....	3
2.2 Systematic Conservation Planning.....	4
2.3 Marxan	7
2.3.1 Input Files	8
2.3.2 Output Files	9
2.4 Species Data.....	10
2.4.1 GIS-Data of Canton Aargau	10
2.4.2 Archive of CCI	10
2.4.3 National Inventory of Dry Meadows and Pastures.....	11
2.4.4 Unique Taxonomic Identifier	11
2.4.5 Red List Status	11
2.5 Cost Calculation.....	12
2.6 Analyses with Marxan	14
2.6.1 Input.dat	14
2.6.2 Spec.dat	14
2.6.3 Pu.dat	15
2.6.4 Puvspr.dat	15
2.6.5 Optimization Runs	16
3. Results.....	18
3.1 Specie Data	18
3.1.1 Species with Red List Status Critically Endangered	20
3.2 Calculation of Current Costs	23
3.3 Best Solutions of the All Species Runs	26

3.4 Best Solutions of the Red List Species Runs	27
3.5 Average Cost per CCI of Best Solutions	28
3.6 Selection Frequencies.....	29
4. Discussion	33
4.1 Species Data Base.....	33
4.2 Cost Calculation.....	33
4.3 Marxan Analysis Results.....	34
4.4 User-defined Targets	35
4.5 Number of Species versus Species Composition	35
4.6 Biodiversity Processes	36
4.7 Dynamics	37
4.8 Transfer Gap between Science and Practice.....	37
5. Conclusions and Future Prospectives	38
Acknowledgment	39
Tables.....	40
Figures.....	41
References	42
Inernet Sources	45
Appendix 1.....	A-1
Appendix 2.....	A-2
Appendix 3.....	A-3

1. Introduction

Taking conservation actions has not arisen recently. It is an old, established practice that has been applied in various cultures all over the world. More recently, conservation areas have been established in order to protect biodiversity and ecosystems. These areas should ensure that conservation features are separated from threats to their existence. But whether this strategy is effective depends on how well it responds to the subsequent two questions. First, how well do conservation areas represent the whole of biodiversity? Second, how well does a reserve systems support the long-term viability of species, ecosystems and the respective essential processes? At this point systematic conservation planning has attended to increase the ability of conservation actions to successfully respond to these questions (Margules and Pressey 2000). The field of systematic conservation planning has emerged over the past 30 years and has influenced planning processes of governmental administrations and non-governmental organisations (Moilanen, Wilson et al. 2009). At the same time, computer based planning tools have been developed and applied worldwide. However, in Switzerland systematic conservation-planning tools have been used only once (Bolliger, Edwards et al. 2011). Hence, this master thesis seek to implement the concept of systematic conservation planning for a part of Switzerland by applying a systematic conservation-planning tool.

The software that is possibly the most widely used systematic conservation-planning tool in the world is Marxan. The objective of Marxan is to solve one type of the minimum set, reserve design problem. Thus, it implements an optimization algorithm that selects planning units based on user-defined targets at minimum cost (McDonnell, Possingham et al. 2002). I apply this optimization algorithm to real data on the species existing in conservation areas of cantonal interest in canton Aargau. My study attempts to illustrate several important issues relevant to implementing a systematic conservation-planning tool for an existing network of conservation areas. First, the study organizes appropriate species data for each of the conservation areas. Second, it includes a method of cost calculation that could be adopted for further analyses with Marxan. Third, it illustrates how Marxan selects the conservation areas with the given cost, the available species data and a variation of user-defined targets. Fourth, it discusses the inputs and outputs of Marxan in considerations of its application as a decision support tool for Swiss conservation

practice. Finally, the study emphasizes the importance of further steps in the planning process in consideration of the key principles of systematic conservation planning.

2. Methods

2.1 Study Site

Canton Aargau is situated in the north of Switzerland and covers approximately 140'400ha in the Swiss lowlands and Jura. The landscape of this canton is diverse. One third of the area is covered with forest and, hence, there are about 4'000km of forest edges. Agriculture land covers about 44% and represents the main type of open land ecosystem. There are about 528km of hedges and tree lines that cross the open land. A small part of the area of canton Aargau, 2.6%, is defined as unproductive land. These are often wetlands that still exist along the rivers Reuss, Limmat and Aare. Canton Aargau has sought to protect such rare habitat types. In fact, it is the canton's obligation to take conservation actions. But this canton's engagement is above the ordinary. Today canton Aargau contains 376 conservation areas of cantonal interest (CCI). Most of these areas are located in open land; only 25 CCIs are in forest (Fig. 1). In the open land, CCIs are dry meadows and pastures, wetlands, bogs, reeds and even former gravel-pits. On the dry meadows and pastures, hedges often exist as well. Especially in the larger CCIs, different ecosystem types and ecological structures are located side by side. The CCIs cover a total area of 2'279ha and, thus, about 1.6% of the cantons area. The smallest CCI constitutes 0.14ha and the largest 109ha (Canton Aargau 2013).

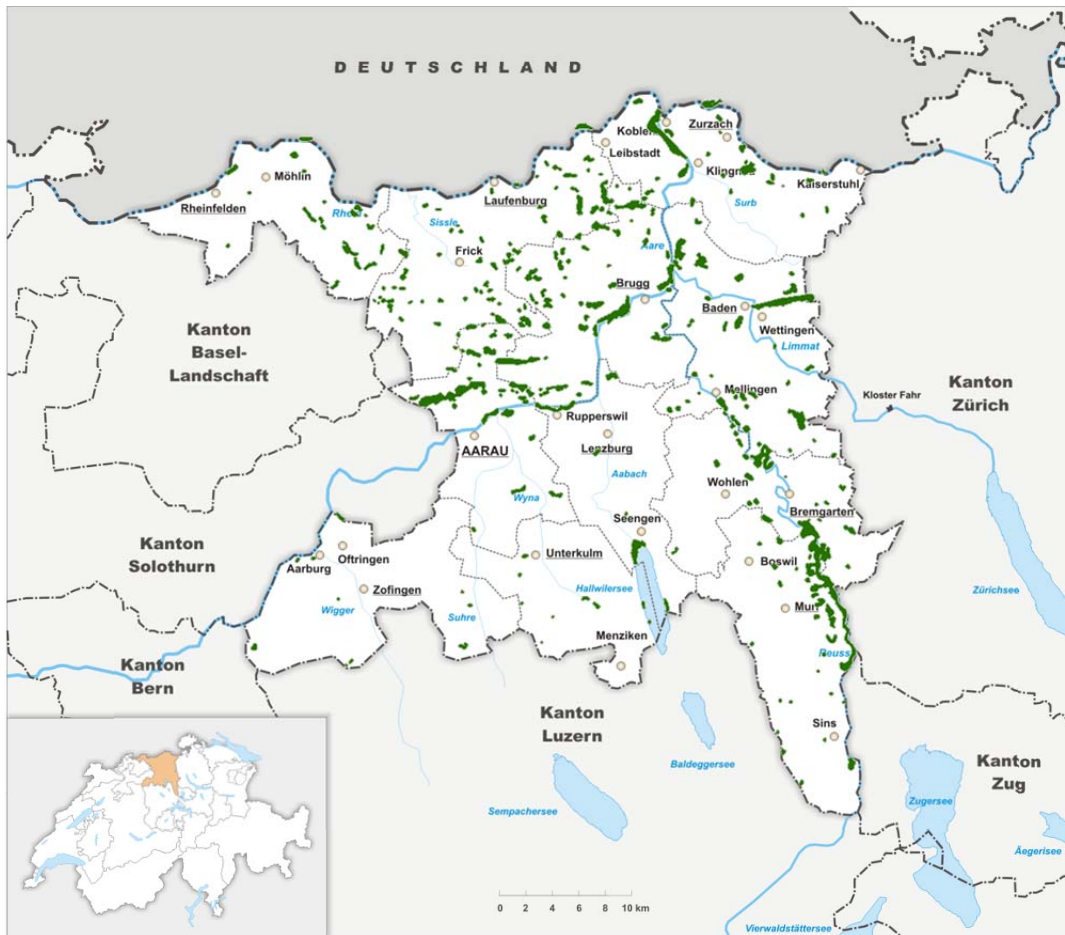


Figure 1: Conservation areas of cantonal interest (green) in canton Aargau

2.2 Systematic Conservation Planning

Taking conservation actions by humans has been an old-established procedure and has taken place worldwide. But protected areas used to be implemented because they were unusable for any commercial purpose or they were abandoned and remote sites. Conservation areas realized in this way don't achieve the two main roles of providing habitat for the biodiversity of each region and break it away from the processes that endangered its survival. The consequences are that all the species which have their habitat in productive areas, are not protected. Thus systematic methods for selecting and designing conservation areas are required (Margules and Pressey 2000).

The field of systematic conservation planning has been evolved for more than 30 years. During this time the term has been defined through several characteristics,

principles and descriptions of the planning process. Margules and Pressey (2000) pointed out six distinctive characteristics of systematic conservation planning:

1. It requires clear choices about the features to be used as surrogates for overall biodiversity in the planning process.
2. It is based on explicit goals, preferably translated into quantitative, operational targets.
3. It recognizes the extent to which conservation goals have been met in existing reserves.
4. It uses simple, explicit methods for locating and designing new reserves to complement existing ones in achieving goals.
5. It applies explicit criteria for implementing conservation action on the ground, especially with respect to the scheduling of protective management when not all candidate areas can be secured at once (usually).
6. It adopts explicit objectives and mechanisms for maintaining the conditions within reserves that are required to foster the persistence of key natural features, together with monitoring of those features and adaptive management as required.

After this, Moilanen, Wilson et al. (2009) crystallized the key principles for spatial prioritization problems (Fig. 2). These principles have arisen with the increasing interest in technical approaches for conservation planning. They all address critical characteristics in the field of planning, designing and managing conservation reserve systems.

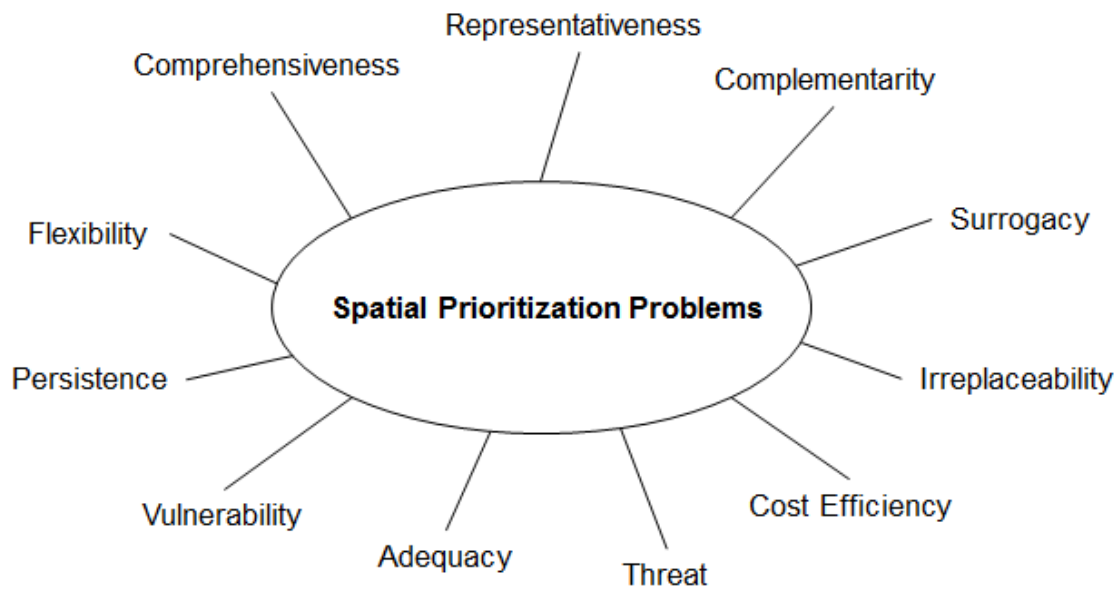


Figure 2: The key principles for spatial prioritization problems have arisen with the increasing interest on technical approaches for conservation planning. They all address critical characteristics in the field of planning, designing and managing conservation reserve systems.

Comprehensiveness and representativeness refer to a network of conservation areas that contains an assemblage of every biodiversity feature including considerations about composition, structure and function. A comprehensive and representative network of conservation areas could be accomplished with a set of complementary conservation areas. To overcome the common lack of data, numerous surrogates have been proposed (Ferrier 2002). The two principles adequacy and persistence point the adequacy of a network for ensuring the permanence of the protected biodiversity features. To comply with these principles considerations on future circumstances have to be taken. Cost efficiency is one principle that systematic conservation-planning tools especially address on. Costs are basic restrictions in taking conservation actions and thus couldn't be discounted. Regarding the target of minimizing biodiversity loss, the respective threats and vulnerability of conservation features have to be incorporated. Further it is crucial to additionally account for future threats. If a planning unit is of particular importance for a reserve system it indicates irreplaceability. This could be the case if a planning unit contains a singular occurrence of a conservation feature. Finally, by allowing for alternative opportunities

in combining planning units, flexibility grants finding a planning unit that has a similar adequacy.

There are several suggestions how to implement the key principles in the planning process. One suggestion of a planning process was the eight stages of systematic conservation planning that has been worked out from several sources (Ardron et al. 2010):

1. Identify and involve stakeholders
2. Identify goals and objectives
3. Compile Data
4. Establish conservation targets and design principles
5. Review existing protected areas and identify network gaps
6. Select new protected areas
7. Implement conservation action
8. Maintain and monitor the protected area network

This study focused especially on the planning stages 3, 4 and 5. Thus, it covered merely a part of the whole long time planning process.

2.3 Marxan

As analytical tool the decision-support software Marxan for systematic conservation planning was applied. Marxan functions to solve the 'minimum set problem', where user-defined targets are met at the lowest possible cost (McDonnell, Possingham et al. 2002, Game and Grantham 2008). In respect to limited resources, it implements the principle of efficiency. To address this principle, Marxan works with different optimization algorithms that implement simulated annealing. Simulated annealing acts by selecting a set of planning units that meet the user-defined targets whereas the score is kept to a minimum (Pearce, Kirk et al. 2008). Targets are set for each species individually. It determines the number of individuals that are intended to exist in the final reserve system. When all the targets are met the score is as high as the sum of the costs of the selected planning unit. If Marxan can't find a reserve system that meets all the targets, the score consists of costs and penalties. The penalty is determined by a species penalty factor (SPF). This factor is a multiplier and could be defined by the user. The penalty is greater the higher the value of the SPF is set. The optimization process takes place several times (determined by the number of runs)

where the current solution is randomly replaced by a more close-by optimal solution. Especially in the beginning of the optimization process with simulated annealing bad moves are stochastically allowed. Through the stochastic way of the simulated annealing optimization algorithm, it avoids getting stuck at local minima (Fig. 3; Ardron et al. 2010). It is obvious that the optimization algorithm would get stuck at the indicated local minimum if the stochastic jumps to the right wouldn't be allowed. This is the key feature of the simulated annealing optimization algorithm in comparison to others.

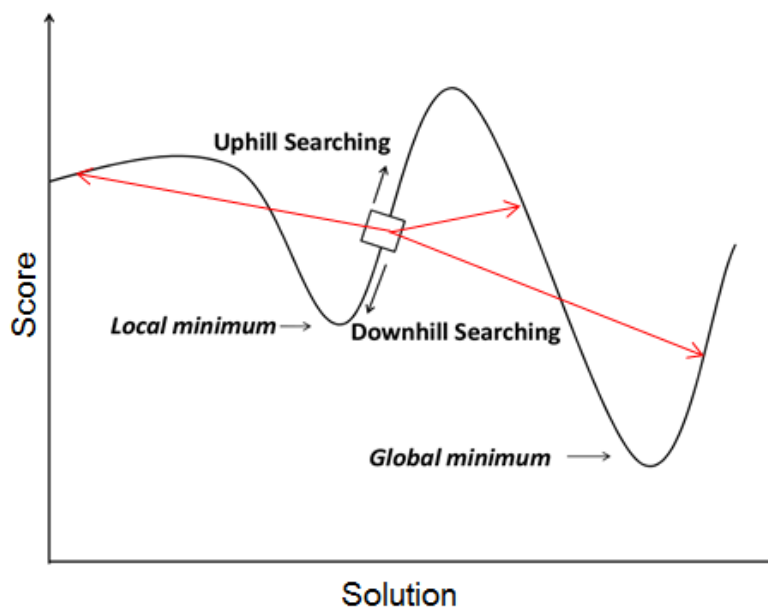


Figure 3: Simulated annealing is an optimization algorithm that could be implemented with Marxan. The main advantage of this algorithm is that it is able to leave local minimum (red arrows). Hence, by allowing these stochastic bad moves it avoids getting stuck at local

2.3.1 Input Files

Marxan requires at least four input files to run an optimization. First, a basic input parameter file (input.dat) is needed. It includes general parameters, the number of iterations, the information on the other input files, the input and output path and further settings that has to be done (Appendix 1). Second, there is the species file (spec.dat) that contains each conservation feature, the corresponding targets and species penalty factor (SPF). The target gives the number of individuals of each species that is intended to be in the solution. If this target is not met, the SPF will be summed up with the cost of the solution to a score. Marxan minimizes this score,

and, thus, the best solution is the one with the lowest score. Third, the planning unit file (pu.dat) is required. This file is a list of the user-defined planning units, their respective costs and their status. The status determines whether a planning unit is locked in or out of the initial and final reserve system (Tab. 1; Game and Grantham 2008). Finally, there has to be a file that connects the conservation feature information with the information on the planning units. This file is named planning unit versus conservation feature file (puvspr.dat).

Table 1: Status value in the planning unit file

Status	Meaning
0	The PU is not guaranteed to be in the initial (or seed) reserve system, however, it still may be. Its chance of being included in the initial reserve system is determined by the 'starting proportion' specified in the Input Parameter File (input.dat)
1	The PU will be included in the initial reserve system but may or may not be in the final solution.
2	The PU is fixed in the reserve system ("locked in"). It starts in the initial reserve system and cannot be removed.
3	The PU is fixed outside the reserve system ("locked out"). It is not included in the initial reserve system and cannot be added.

2.3.2 Output Files

Marxan produces several outputfiles that illustrate the results in different ways. There is one separate output file for each iteration if the same run is repeated several times. These files contain a list of all planning units in the first column. In the second column, there is either a "0" for not included or a "1" for included in the solution. Among these files, there is the best solution file. It contains the same as the others but for the best solution in the series of the repeat runs. The summary information file gives an overview of the iterations of one run. There the results of each iteration could be compared in consideration of the score, the cost, the number of planning units, the number of missing values (here species) and the penalty costs of the found solution.

2.4 Species Data

2.4.1 GIS-Data of Canton Aargau

The online Geoportal of canton Aargau provided different GIS data of the canton. I used five GIS-layers in the course of the analyses (Tab. 2). First, the amphibian inventory layer contained spatial point data of frog species, toad species, newt species and one salamander species records. Second, the layer “Fledermausquartiere” contains recorded nests of several bat species. Third, the CCIs of canton Aargau are digitized in a polygon layer with additional information on the municipality name, the field name, the habitat type and the size. As the amphibian-inventory, the ornithologic and the reptilian inventory contained point data of recorded species. There were duck and bird species in the ornithologic inventory. The reptilian inventory contained saurian and snake species.

Table 2: Used GIS-layers of canton Aargau

Description	Name	Type	Investigation Date	Source
Amphibieninventar 91/92	AGIS.alg_amphibien92	Vector (point)	1.1.1991 to 1.12.1992	© Aargauisches Geografisches Informationssystem (AGIS)
Fledermausquartiere	AGIS.alg_fledermausquar	Vector (point)	1973 to 1993	© Aargauisches Geografisches Informationssystem (AGIS)
Naturschutzgebiete v.kant.Bed. 1:5'000	AGIS.alg_natschgebkb5t	Vector (polygon)	1.1.1993 to 1.12.1996	© Aargauisches Geografisches Informationssystem (AGIS)
Ornithologisches Inventar	AGIS.alg_ornithinv	Vector (point)	1.1.1985 to 1.12.1987	© Aargauisches Geografisches Informationssystem (AGIS)
Reptilieninventar: erfasste Arten	AGIS.alg_reptinvea	Vector (point)	1.1.1987 to 1.1.1990	© Aargauisches Geografisches Informationssystem (AGIS)

2.4.2 Archive of CCI

A great deal of the data were out of the CCI archive of Canton Aargau. This archive consisted of non-digitized inventory data taken from surveys of conservation areas ordered by the municipalities. The first step was to find if out whether these inventory data belonged to a CCI and if yes to which one. Some inventory sheets were marked with the CCI number; others were allocated by coordinates, field name or map in comparison with the GIS-layers from canton Aargau or swisstopo (Tab. 2, 3).

Table 3: GIS-layer that contained field names

Description	Name	Type	Investigation Date	Source
Pixelkarte	pk25_komb	Raster	2013	© swisstopo

2.4.3 National Inventory of Dry Meadows and Pastures

Another source of species data, mainly for plant species data, was the national inventory of dry meadows and pastures (Trockenwiesen- und weiden von nationaler Bedeutung BAFU, Data contract 31.10.2013). It contained plant species records in all the CCIs that are in the national inventory as well. The records were derived from the years 1997 and 1998.

2.4.4 Unique Taxonomic Identifier

To add a unique taxonomic identifier (ID) to every plant species, the Landolt-number (Landolt 1977) extended because some inventory sheets of the CCI archive already contained it for each recorded species. All other Landolt-numbers could be found in the online database of the national data and information center of the Swiss flora (infoflora.ch 2013).

The taxonomic identifiers from the Fauna Europaea (de Jong 2013) were adopted for all the animal species. They didn't overlap with the plant IDs and could be found online for additive records.

2.4.5 Red List Status

Based on the Swiss red lists for animals and plants (Keller et al. 2010, Monnerat et al. 2007, Moser et al. 2002, BAFU 2013), the red list status was added to each recorded species by means of the scientific name. The red list status of each species that was given in values from 0 to 4 categories before 1994 were cross-walked to the new system that is compatible with that of the IUCN (Tab. 4). The species that had the status extinct, critically endangered, vulnerable or near threatened were considered as "red list species".

Table 4: Categories of red list status (source: BAFU)

Categories before 1994	Valid Categories	
0	EX	Extinct
0	EW	Extinct in the Wild
1	CR	Critically Endangered
2	EN	Endangered
3	VU	Vulnerable
4a-4d	NT	Near threatened
n	LC	Least Concern
-	DD	Data Deficient
-	NE	Not Evaluated

2.5 Cost Calculation

Only the current costs were calculated for each CCI since there was no information on other types of costs. Current costs were defined as costs for relinquishment of management, compensation money and costs for maintenance. Costs for contracts, planning, acquisition and implementation of new conservation areas were not considered. I based the cost calculation on the report of Ismail et al. (2009). After that, maintenance costs depend on ecosystem type (forest, pasture, meadow, bog), slope and patch size. There are minimum and maximum basic costs per ha and year that depend on ecosystem type. The minimum basic amounts were chosen concerning the optimization problem in Marxan (Tab. 5). For the open, land there was a surcharge per hectare and year depending on the slope of the terrain of the conservation area (Tab. 6). Additionally, there was a surcharge or a reduction per hectare and year for open land depending on the size of the parcel (Tab. 7). Finally, the cost for maintenance of hedges was calculated by considering the value of the direct payments for hedges in Switzerland per hectare and year (ALN Zurich 2013). The length of hedges per CCI was derived from the GIS layer TLM_Baum_Gebüschreihe_2012 (Tab. 8). I multiplied the total length per CCI with a medium width of four meters (ALN Zurich 2013). The total hedge area per CCI multiplied with the 2500 Swiss francs direct payments per hectare hedges resulted in the total cost for the maintenance of the hedges per CCI. This cost was finally added to the sum of the ecosystem type costs per CCI.

Table 5: Basic costs per 1ha per year

Ecosystem type	Swiss Francs
Forest	190.00
Pasture	827.00
Meadow	1443.00
Bog	1517.00

Table 6: Surcharge per hectare and year depending on the slope

Slope	0-18%	18-35%	35-50%	50-80%	>80%
Pasture	3.50	15.00	28.00	45.00	45.00
Meadow	27.00	114.00	170.50	317.50	1810.50
Bog	30.00	156.50	205.00	457.00*	2296.00*

*only virtual, no such case

Table 7: Surcharge or reduction per hectare and year depending on the patch size

Patch Size	<0.5ha	0.5-1.0ha	1.0-2.0ha	2.0-5.0ha	>5.0ha
Pasture	21.00	11.00	0.00	-6.00	-8.00
Meadow	248.50	114.00	0.00	-104.00	-212.00
Bog	282.00	141.00	0.00	-155.00	-255.50

The CCI polygons were transferred to a raster of 25m to 25m size since there were CCIs including different ecosystem types and slopes,. This raster was overlaid with the land cover type layer and the digital elevation model layer (Tab. 8). As a result, each CCI was split in several pixels, which contained the information of ecosystem type and slope. As the land cover type layer does not include the three categories pasture, meadow and bog for open land, the description of the ecosystem type per CCI in the attribute table of the CCI layer defined the category. I calculated parcel size by summing up all pixels of the same ecosystem type. The total cost of each CCI was calculated by multiplying the area with the corresponding basic cost, slope surcharge and size surcharge or reduction(Tab. 5, 6, 7).

Table 8: Used GIS-layers for cost calculation

Description	Name	Type	Year	Source
Primärflächen	geolib.pri25_p_07	Polygon	2007	© swisstopo
Geländedaten: DHM25	geolib.DHM25	Raster	1994	© 1995 Bundesamt für Landestopograp hie
TLM_Baum_Gebueschreih_2012	geolib.swissTLM3D_1_1_BB	Line	2012	© swisstopo

2.6 Analyses with Marxan

I conducted several Marxan runs with a variety of settings in order to find out how Marxan works with real data of the CCIs. Of course the focus was on the question how many CCIs are required to meet the given targets at minimum cost. On one hand I conducted several runs where targets were set for all species equally (“all species runs”) other runs in which only the red list species were targeted (“red list species runs”). In these two groups the parameters “target” in the input file spec.dat and “amount” in the input file puvspr.dat have basically been changed. Because the collected data was often presence data, the numbers of individuals of a species in a CCI wasn’t known. So first the amount was set for all species to the same values (1, 2, 5 and 10). Second, random values ranging from 1 to 10 were produced to make the data set more realistic (random.org 2014). So, the amount value for each species varied. The remaining parameters in all the input files stayed the same for every run in this study. Thus, the results of very similar runs allow comparability and may illustrate how the optimization algorithm of Marxan works with the real data of canton Aargau. However, the pragmatically defined and simple targets were used to do this experiment with hypothetical goals. Furthermore, by ascending the targets limits could be found where targets could not be met.

In the following subchapters the four input files are described in detail with regard to the data of this study. The different input files were produced with R-Studio the editor for the statistic software R except the input parameter file.

2.6.1 *Input.dat*

The applied input parameter files were produced with the included file maker inedit.exe. I mainly used the default Marxan parameters as recommended in the manual (Appendix 1). I only changed the number of repeat runs (NUMREPS) from “1” to “10”. Marxan conducted ten runs instead of one run for each parameter setting. Thus, the probability of finding a near optimal solution was higher. In addition, I conducted all runs with simulated annealing by setting the runmode (RUNMODE) to “1”.

2.6.2 *Spec.dat*

The three column names of the species input file need to be “id”, “target” and “spf”. The column “id” should be filled with unique identifiers of each conservation feature

(Tab. 9). There were several possible types of conservation features. After Ardron et al. (2010), a conservation feature was defined as an element that the user would like to occur in the final reserve system. So, I defined it as species that could potentially occur in the final reserve system. Depending on the optimization problem, various targets were added to the species. The SPF was set to “2” in all runs.

2.6.3 *Pu.dat*

One of the first steps before running Marxan was to define the planning units. Based on the collected data, the CCIs of canton Aargau suggested itself to operate as planning units. So, in this study the 376 CCIs were implemented as planning units whereupon the official IDs were adopted. The cost of each CCI was then calculated as it was described in chapter 2.5. The values were rounded and added to the respective CCI in the planning unit file (Tab. 10). The status as it was described in table 8 was set to “0” for all CCIs in every run. The complete list of the CCIs and their respective cost was added to the appendix 1.

2.6.4 *Puvspr.dat*

The planning unit versus conservation feature file contained a list with the information of which conservation feature occurred in which planning unit. In the first column the conservation feature IDs were listed as many times as they occur in different planning units. In the second column the planning unit IDs were arranged next to the conservation feature they include (Tab. 11). So each planning unit was listed as many times as they contain conservation features. The information of how many of the respective conservation feature existed in a particular planning unit, was listed in the third and last column “amount”. For this study every unique record of all plant and animal species was listed. As mentioned the amount was changed depending on the conducted run.

Table 9: spec.dat

id	target	spf
8	1	2
14	1	2
18	1	2
21	1	2
22	1	2
27	1	2
...

Table 10: pu.dat

id	cost	status
1	4357	0
2	636	0
3	1942	0
4	1477	0
5	10'102	0
6	10'702	0
...

Table 11: puvspr.dat

species	pu	amount
8	330	1
8	362	1
14	159	1
18	25	1
18	162	1
21	28	1
...

2.6.5 Optimization Runs

In table 12 the target and amount of each run was listed. The runs were named as indicator for the settings. So, for example “T1A1” means that the target was set to “1” (T1) and the amount was set to “1” (A1). The target value “1” meant that one individual of each species should occur in the final reserve system. The amount value stood for the number of individuals per CCI of each species. In several runs the random values (R) were set for the amount column. When only the red list species were targeted, “RL” was added to the run name. Assuming the ratio of the target and the fix amount were critical to the best solution different runs with the same ratio were conducted (e.g. T1A1, T2A2, T5A5 with ratio 1). Because the amount is at least as high as the target, I expected similar best solutions for runs T1A2 and T1AR. For these reasons there were eight unique runs. These unique runs were highlighted in bold face in table 12.

Table 12: Optimization Runs where the targets are the same for all species; R means random amounts

Run	Target for not threatened species	Target for red list species	Amount
T1A1	1	1	1
T1A2	1	1	2
T2A1	2	2	1
T2A2	2	2	2
T5A1	5	5	1
T5A2	5	5	2
T5A5	5	5	5
T10A1	10	10	1
T10A2	10	10	2
T10A5	10	10	5
T1AR	1	1	R
T2AR	2	2	R
T5AR	5	5	R
T10AR	10	10	R

I conducted another seven runs in which only the red list species were targeted (Tab. 13). Since these species were not found in numerous CCIs the target was only set to “10” when implementing the random amounts. These random amounts were adopted from the runs above and thus stayed the same for all the runs with random amounts.

Table 13: Red list species runs. R stands for random amounts

Run	Target for not threatened species	Target for red list species	Amount
RLT1A1	0	1	1
RLT2A1	0	2	1
RLT5A1	0	5	1
RLT5A2	0	5	2
RLT2AR	0	2	R
RLT5AR	0	5	R
RLT10AR	0	10	R

3. Results

3.1 Specie Data

Merging the data of the different sources resulted in 17'460 records in 339 CCIs. These records minus the double counted due to different record years then turned to 14'329 records. They consisted of 832 plant species and 369 animal species, i.e. 1201 species in total. For 37 CCIs no species data were available. In total there were 1729 records of red list species in 278 CCIs (Tab. 14). These records consisted of 714 plant species records and 1015 animal species records. These records derived from 97 red list plant species and 99 red list animal species.

Table 14: Summary information red list species data

Records	
Number of records of total red list species	1729
• Number of records of red list plant species	714
• Number of records of red list animal species	1015
Species	
Number of total red list species	196
• Number of red list plant species	97
Status CR – critically endangered	2
Status EN – endangered	24
Status VU – vulnerable	71
• Number of red list animal species	99
Status CR – critically endangered	9
Status EN – endangered	27
Status VU – vulnerable	63
Conservation Areas	
Number of CCIs with red list species	277

The CCI with the number 88 had the maximum species number in both cases (Figs. 4, 5; highlighted in light blue). These were in total 360 species including 45 red list species. Thereby it was the CCI with the most recorded species by far. With 241 and 222 species in total and accordingly 23 and 18 red list species, the CCIs with the number 362 and 159 respectively were on the second and third position in the species ranking (in red in Figs. 4 and 5).

In both of the two figures the category with the lowest numbers of species (green) was the one with the most CCIs within. Next to the 37 CCIs that had no species data at all there were 223 CCIs that had 1 to 51 species in total. Regarding the red list

species there were another 62 CCIs that had no species records. For 184 CCIs 1 to 6 red list species were recorded.

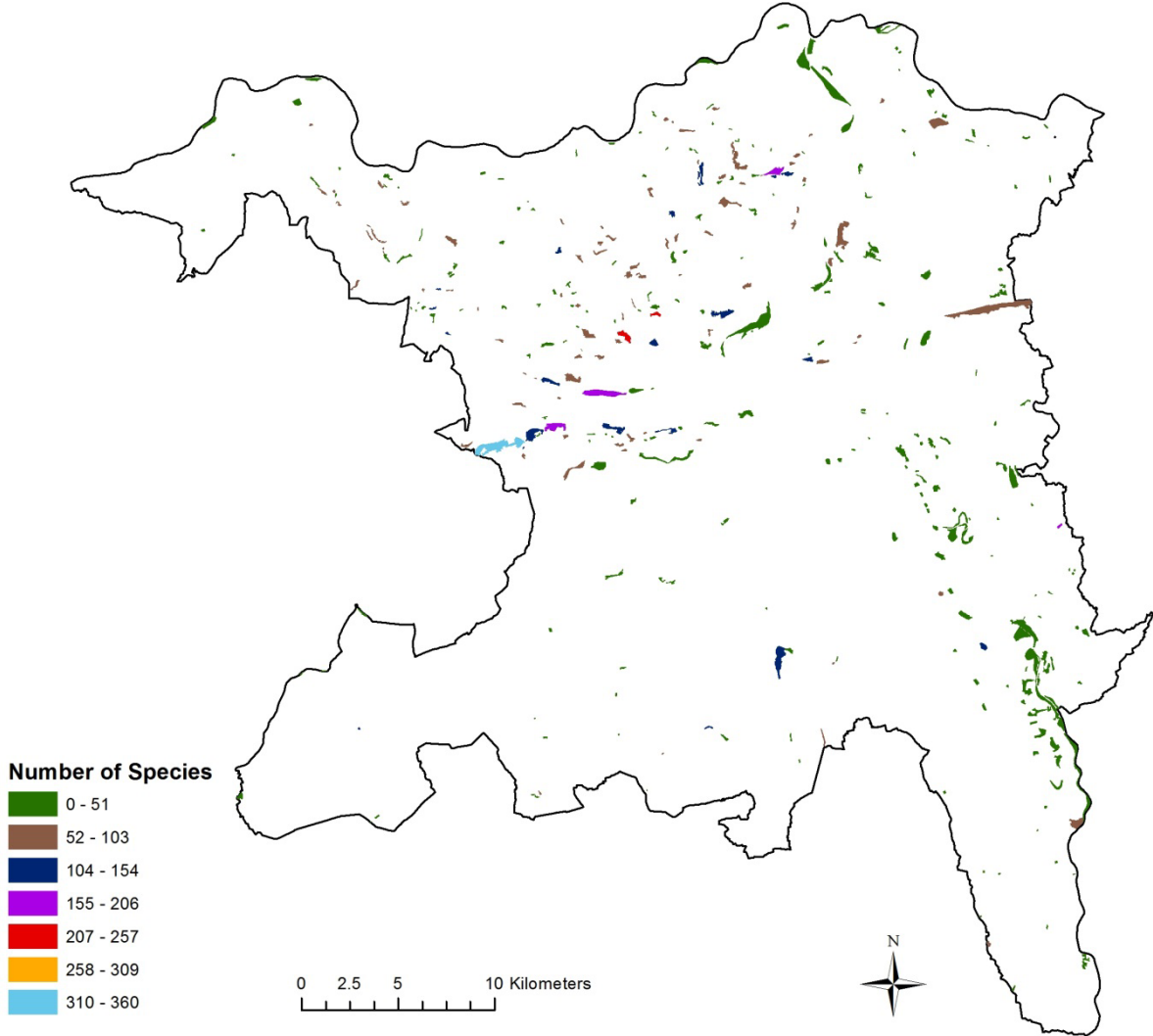


Figure 4: The number of species per CCI. The CCI 88 in light blue was the one with the most recorded species. In contrary, there were no data available for 37 CCIs. For the main part, 223 CCIs, 1 to 51 species were recorded.

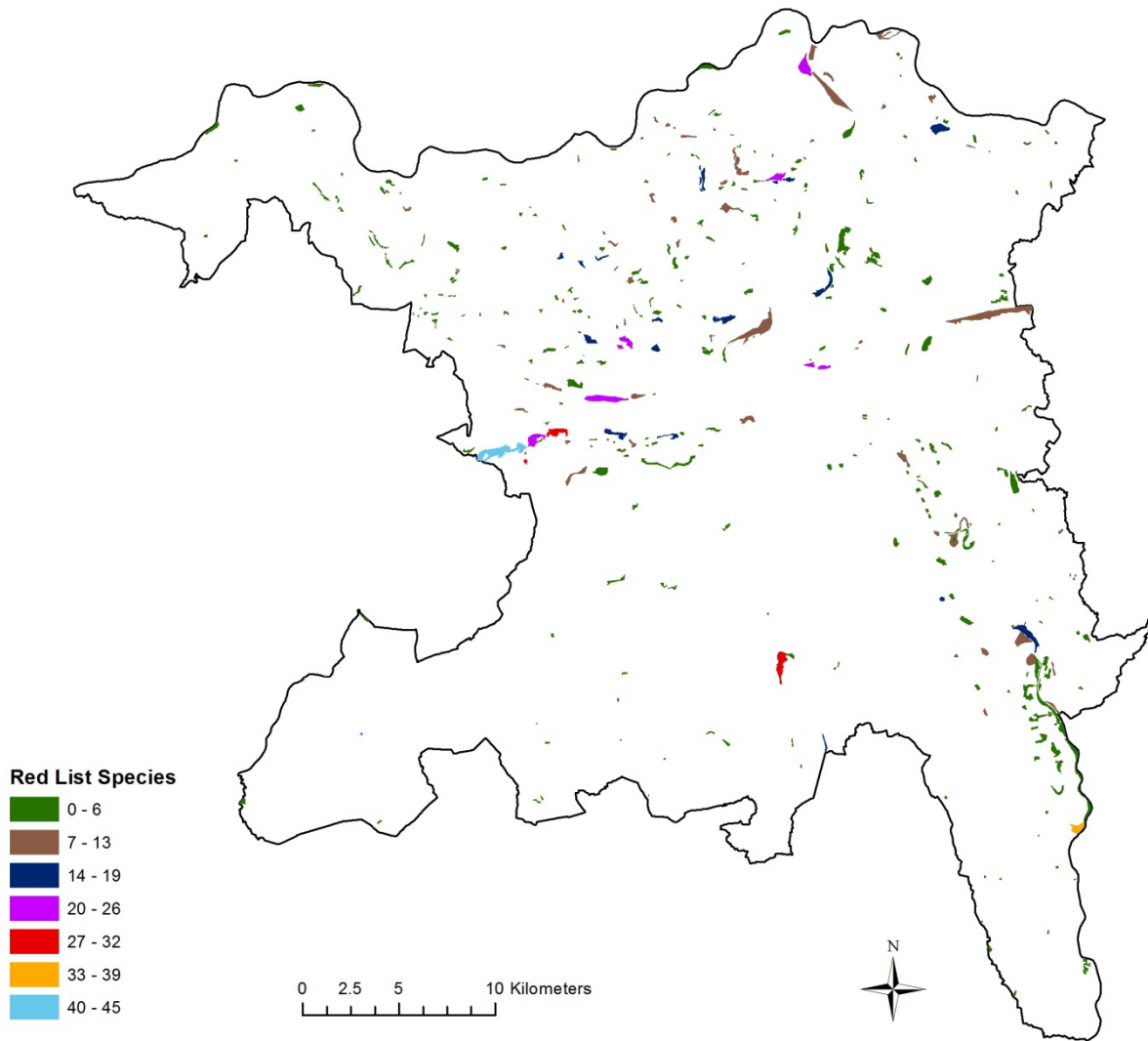


Figure 5: The number of red list species per CCI is illustrated in seven equal intervals. Here, even for 99 CCIs no data were available. For another 148 CCIs only 1 to 6 red list species were recorded. Again, the CCI 88 had the maximum number of red list species.

3.1.1 Species with Red List Status Critically Endangered

There were 11 species with red list status CR (critically endangered) in the collected species data (Fig.6-16). Two of them were plant species and nine were animal species. The plant species were both phanerogams (*Sium latifolium*, *Dactylorhiza maculata*). The animal species consisted of one snake species (*Vipera aspis*), two butterfly species (*Satyrrium pruni*, *Cupido argiades*), three dragonfly species (*Leucorrhinia pectoralis*, *Leucorrhinia caudalis*, *Sympetrum pedemontanum*), one mammalian species (*Castor fiber*) and two bird species (*Lanius senator*, *Vanellus vanellus*).



Figure 6: *Sium latifolium*



Figure 7: *Dactylorhiza maculata*



Figure 8: *Vipera aspis*



Figure 9: *Satyrium pruni*



Figure 10: *Cupido argiades*



Figure 11: *Leucorrhinia pectoralis*



Figure 12: *Leucorrhinia caudalis*



Figure 13: *Sympetrum pedemontanum*

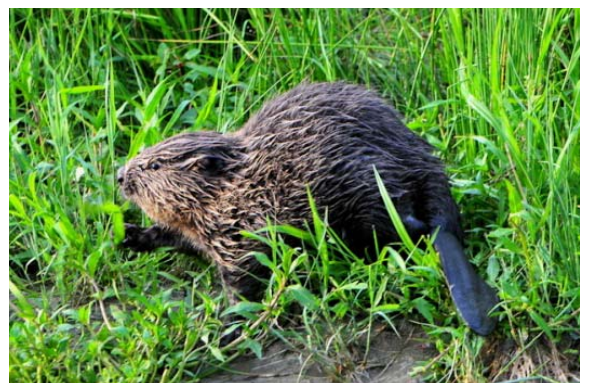


Figure 14: *Castor fiber*



Figure 15: *Lanius senator*



Figure 16: *Vanellus vanellus*

3.2 Calculation of Current Costs

The current cost per year for each CCI added up at maximum 54'620 Swiss francs. This maximum cost was reached by the CCI with the number 60 (highlighted in green in Fig. 17). This CCI is the well-known Boniswiler Ried at the lake of Hallwil. It is the largest remaining low moor in canton Aargau. By far it was the most expensive CCI, so there was no CCI within the next lower category (brown). Thereafter, only one CCI was in the next lower category (dark blue), number 88 the same CCI with the most recorded species. The costs for another two CCIs reached the medium level category (pink). They are located in the south east of the canton at the riverside of Reuss. The categories highlighted in red and orange covered 11 and 35 CCIs respectively. Finally, for 326 CCIs and thus for the main part of the CCIs, the current cost was estimated not to be higher than about 7800 Swiss francs per year (light blue). The CCI with the least cost was number 62 that consisted of a pond. A detailed list of the current cost for each CCI could be found in the appendix 1.

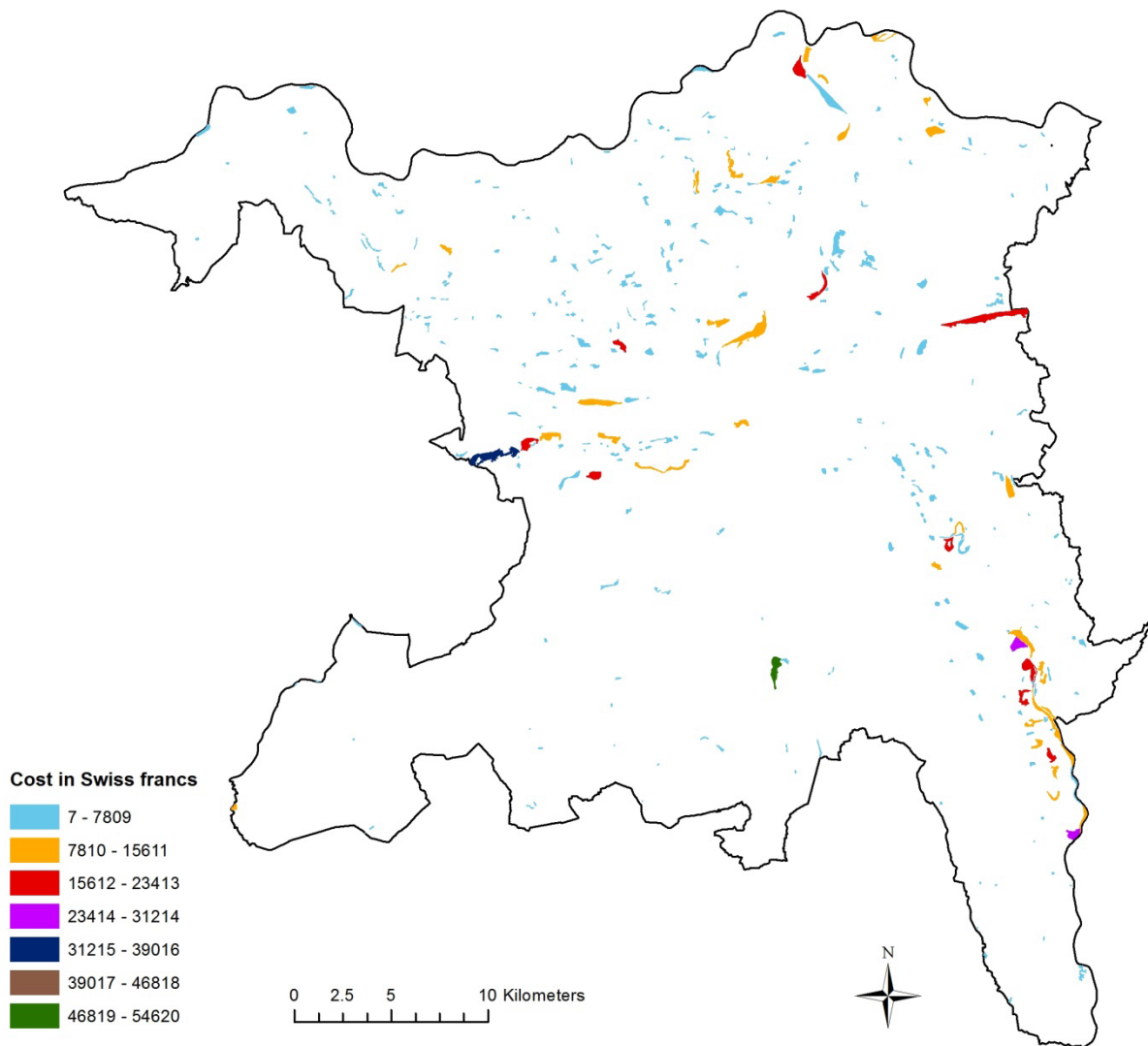


Figure 17: The current costs per year for each CCI was calculated and illustrated in seven equal categories. In general, these costs were not higher than 10'000 Swiss francs. There were some exceptions as the most expensive CCI, the Boniswiler Ried at Lake Hallwil (green).

To bring the species data together with the cost data, each dot in figure 18 represented one of the 376 CCIs. In general the CCIs had a total number of species within 0 and 100 at a yearly cost of up to 20'000 Swiss francs. There were only a few outliers. On one hand there were the CCIs with number 88, 362 and 159 which had more than 200 species. On the other hand there were the expensive CCIs as the number 60 and again 88. The figure 18 illustrated that there were a couple of species-rich CCIs that had yearly costs less than 10'000 Swiss francs. Even though there were CCIs that had yearly costs higher than 10'000 Swiss francs but had not more than 50 recorded species. As the current cost per CCI was calculated per hectare they were expected to correlate with the respective size of a CCI. The plotted data in figure 19 showed this correlation. Thus, larger CCIs had higher total costs.

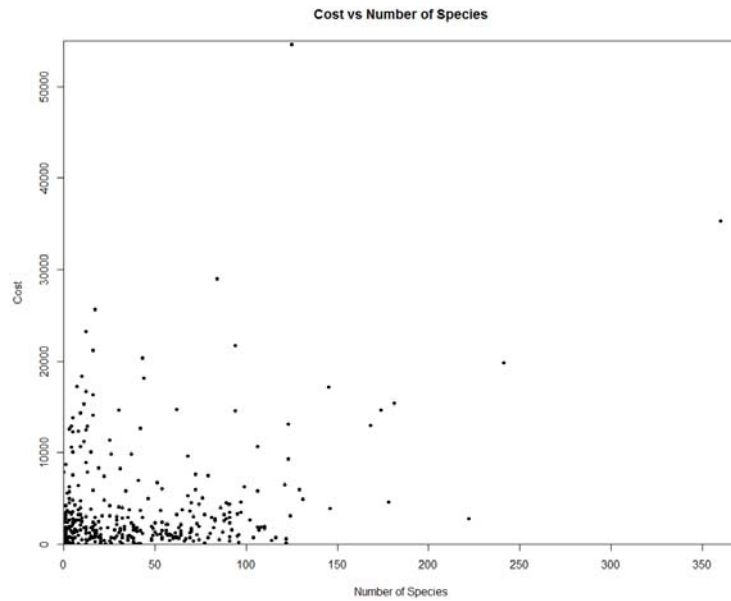


Figure 18: The calculated costs were plotted against the number of recorded species in each CCI. The most CCIs had a number of species from 0 to 100 and yearly current costs in between 0 and 10'000.

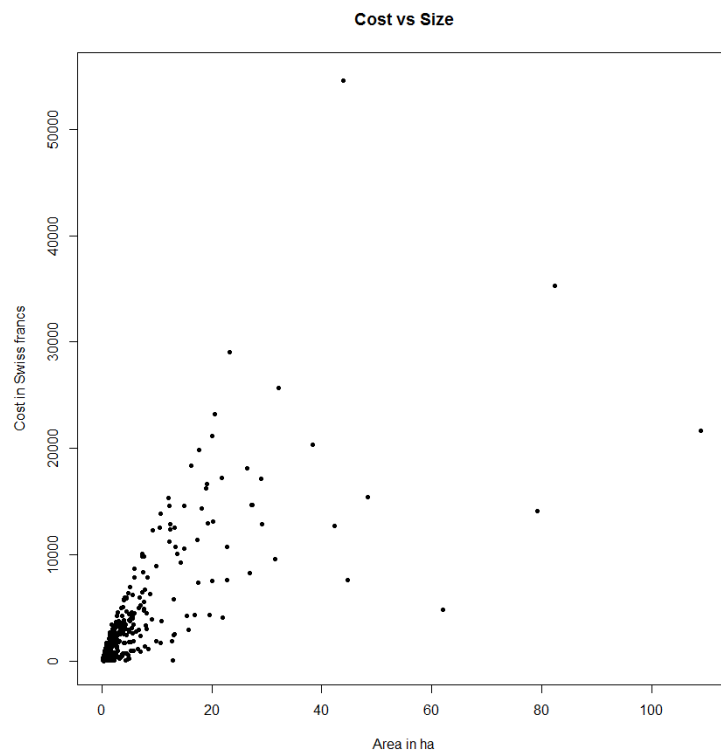


Figure 19: The calculated cost of each CCI was plotted against the respective size in hectares. As the cost was calculated per hectare and then summed up for each CCI, it was expected to correlate with the size.

3.3 Best Solutions of the All Species Runs

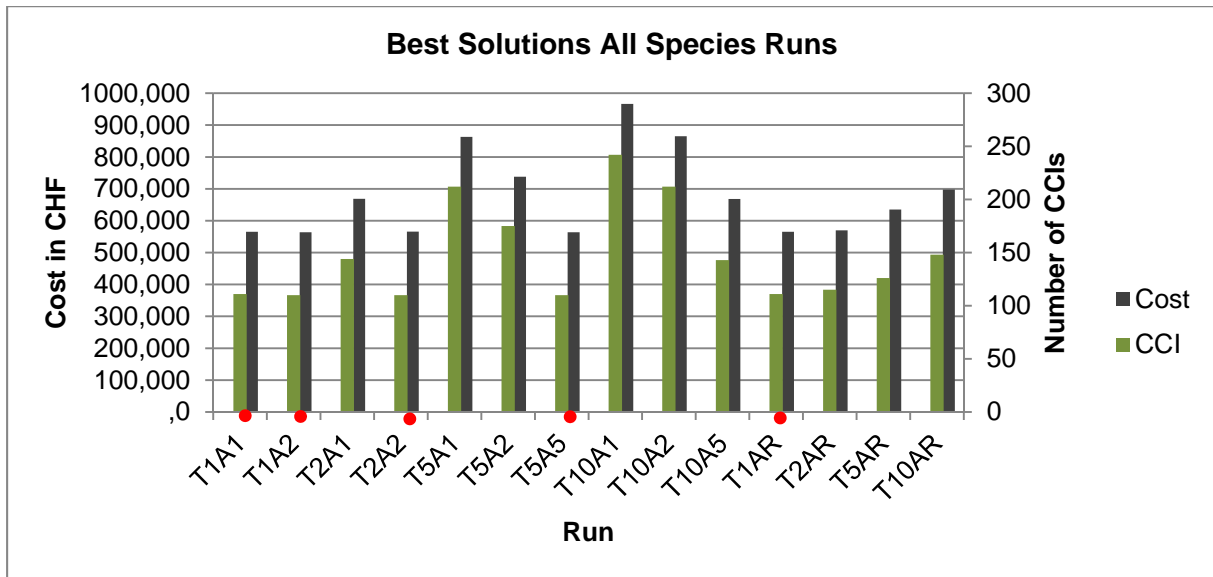


Figure 20: The cost and the number of selected CCIs of the respective best solution of each run was compared. The higher the targets and the lower the amount the higher were the costs and the number of CCIs for a optimal reserve system.

Targets could be met in five of the 14 optimization runs (see red dots in Fig. 20). Not all of the species were involved in the best solution of a final reserve system in the remaining nine runs. As expected the best solutions were mostly the same when the ratio of target and amount was the same. Following groups emerged to have the same or nearly the same best solution (Tab. 15).

Table 15: Similar runs grouped

Group 1	Group 2	Group 3
T1A1	T2A1	T5A1
T1A2	T10A5	T10A2
T2A2		
T5A5		
T1AR		

In group 1, with target-amount ratio of 1 or below, the targets could be met. When only the eight unique ratios were considered, the targets could be met for runs where the amount was at least as high as the targets (e.g. T1A1, T1A2). Targets were not met when the amount was lower than the target (e.g. T2A1, T5,A1). If each of these groups (Tab. 15) were seen as one run, targets could be met in only one of the eight unique runs. In group 2, targets were not met for 390 species. In group 3, targets

were not met for almost 60% of the species (711). In fact in run T10A1 for even 890 species targets were not met (Tab. 16).

Table 16: Number of missing species in the best solution

Run	Missing Species
Group 1	0
T2AR	29
T5AR	162
Group 2	390
T10AR	426
T5A2	549
Group 3	711
T10A1	890

Overall with the lowest demands (T1) at least 110 of the CCIs were required (group 1). The best solutions of the runs T2AR, T5AR and T10AR involved 115, 127 and 148 CCIs respectively. In the runs of group 2 and 3, 144 CCIs and 212 CCIs were chosen, respectively. For the best solution with the settings of run T5A2 and T10A1 Marxan computed 175 and 242 CCIs respectively. As expected, the best solutions spanned more CCIs the higher the targets and the lower the amounts (Fig. 20).

The respective cost of the best solutions ranged from 564'037 CHF (in T1A2) to 174'163'829 CHF (in T10A1) at which the cost was higher when the targets were not met. In general, the higher the ratio of target to amount, the higher is the risk of not meeting the targets and, thus, the higher the cost of the best solution (e.g. T10A1). This is due to the fact that more CCIs are needed the higher the amount is.

3.4 Best Solutions of the Red List Species Runs

There was one best solution among the seven red list species runs that met all the targets (Fig. 21). It was the run with the lowest target RLT1A1. In the remaining best solutions at least six and maximum 132 species were not involved (Tab. 17). This maximum of missing species was about 65% of the total red list species. In general, the same characteristics as above could be seen in the selection of CCIs for the best solution. Targets were only met when the amount was at least as high as the targets. But since there were less species the best solutions absolutely involved less CCIs. In fact Marxan chose 48 CCIs in the best solution of the run RLT1A1 which had the lowest target. An additional two CCIs were included in the best solution of run

RLT2AR. Further, the best solutions of the runs RLT5AR, RLT10AR and RLT2A1 consisted of 58, 71 and 72 CCIs respectively. Finally, the best solution with the most missing species was the one that contained the most CCIs.

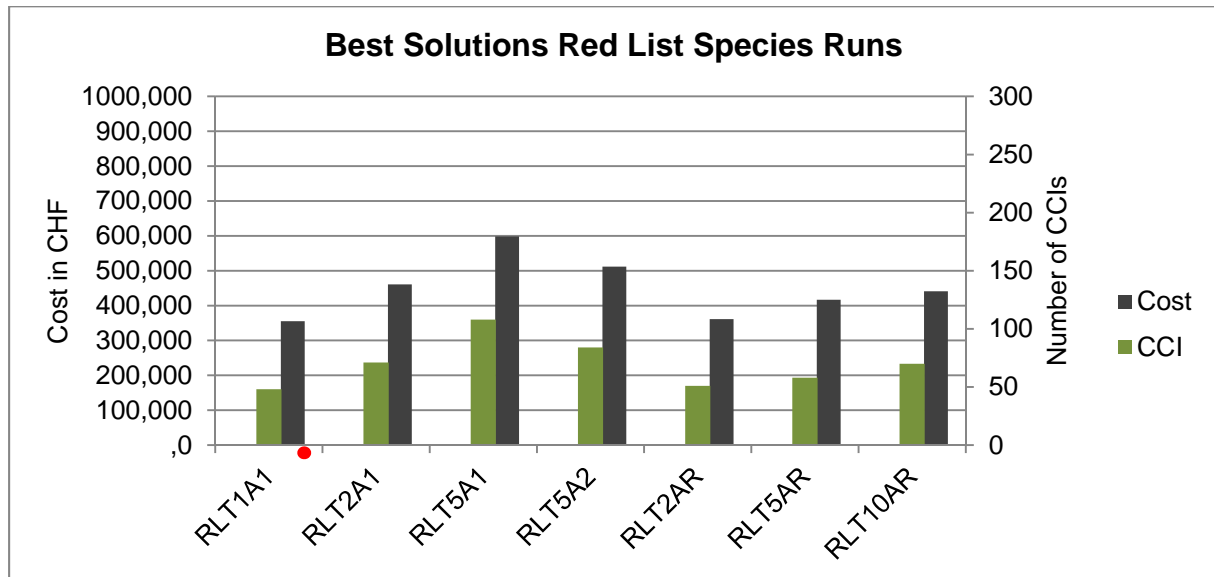


Figure 21: The costs and number of CCIs of the best solutions of the red list species runs were plotted in bars. The number of CCIs in a solution ranged from 44 to 108; the costs from 355'321 to 598'772 Swiss francs.

Table 17: Number of missing species in the best solutions of the red list species runs

Run	Missing Species
RLT1A1	0
RLT2AR	6
RLT5AR	30
RLT2A1	73
RLT10AR	90
RLT5A2	110
RLT5A1	132

3.5 Average Cost per CCI of Best Solutions

In comparison to the best solutions with all the species involved, the cost per CCI was higher in the best solutions with the red list species. Even the lowest average cost in the best solution of run RLT5A1 was higher than the highest average cost of the runs in group 1 (Fig. 22). In both cases the average cost emerged in the best solution where all targets could be met (group 1 and RLT1A1). These two solutions involved the lowest number of CCIs. This indicates that Marxan chose fewer, but more expensive, CCIs to meet the given targets.

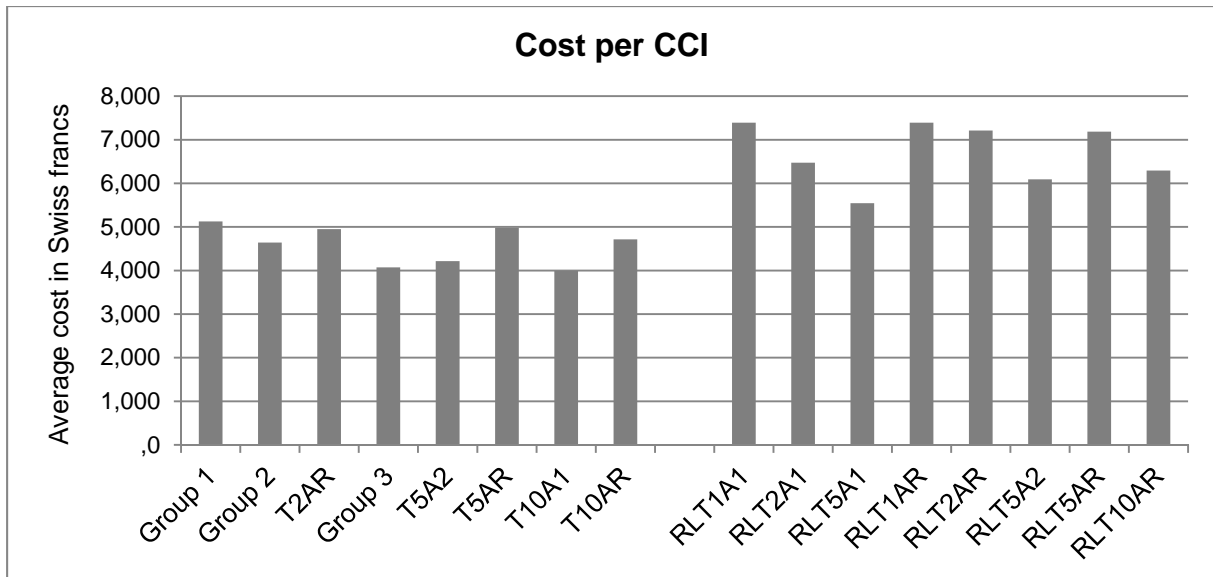


Figure 22: The average cost per CCI in the respective best solution of each run was calculated and compared. The result showed that even the lowest average cost in the best solution of run RLT5A1 was higher than the highest average cost of the runs in group 1.

3.6 Selection Frequencies

As another result, the selection frequency of each CCI over the best solutions of the eight unique all species runs was calculated (Fig. 23). This frequency ranged from 0 meaning never selected in a best solution to 8 meaning selected in all the best solutions. The colors corresponded to the seven equal categories of this frequency and correspond to the colors used in the maps in chapter 3.1 and 3.2. The colors in this map illustrates that a big part of the CCIs was never been selected in these eight runs (green). Contrarily another large part was involved in every best solution (light blue). About 134 CCIs were never selected while 105 CCIs were always included in the best solution (Fig. 24). Seven to 33 CCIs had a selection frequency in between 0 and 8.

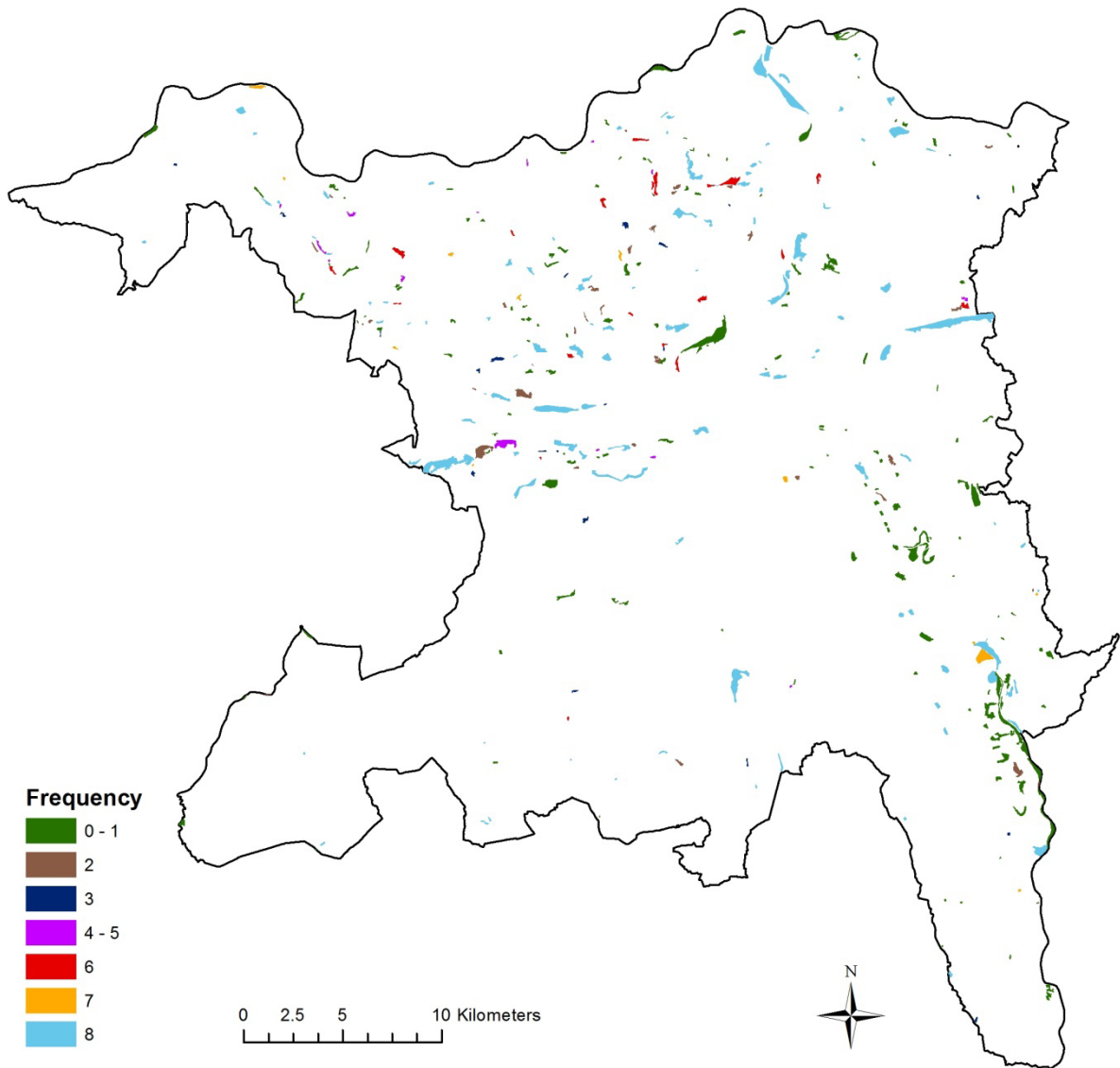


Figure 23: The selection frequency of each CCI over the eight unique all species runs was illustrated. 105 CCIs were included in every best solution. In contrast, another 134 CCIs were never selected.

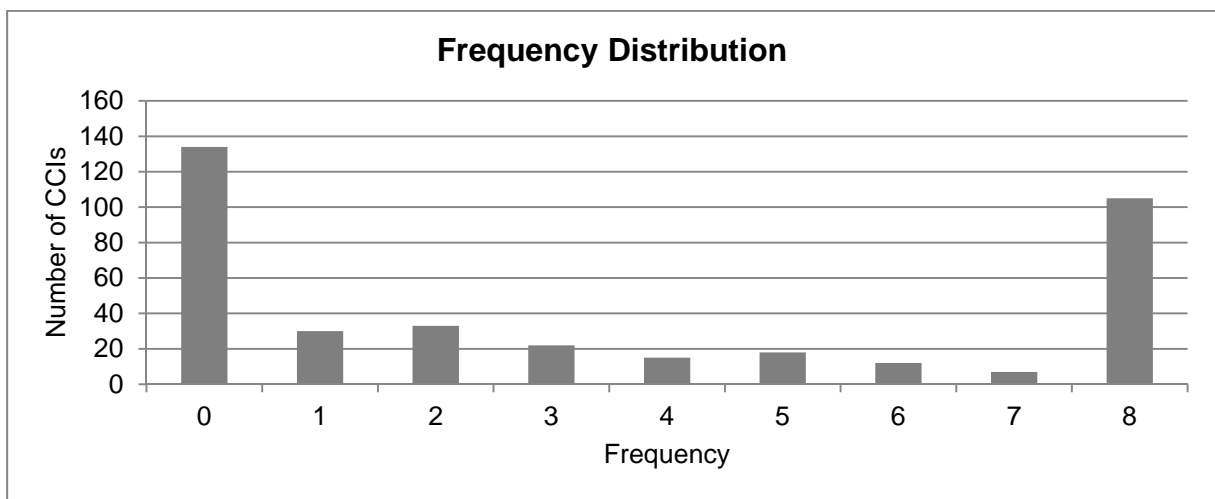


Figure 24: The Number of CCIs was plotted as a bar for each frequency. This frequency distribution illustrated that a CCI was either selected never or in every best solution.

The same as above was done for the red list species runs. A similar result as above emerged (Fig. 25). There were a lot CCIs with a selection frequency either in the lowest (green) or in the highest (light blue) category. But this time the number of CCIs that were never involved in a best solution, was twice as high as with the all species runs (Fig. 26). This time, even 268 CCIs were never selected in a best solution. Contrarily the number of CCIs that were always selected was quite low. Only 44 CCIs were involved in every best solution. Each of these 44 CCIs was in the highest selection frequency category from above too. If the two maps were compared, especially the large CCIs in light blue attracted attention (see red circles in Fig. 25).

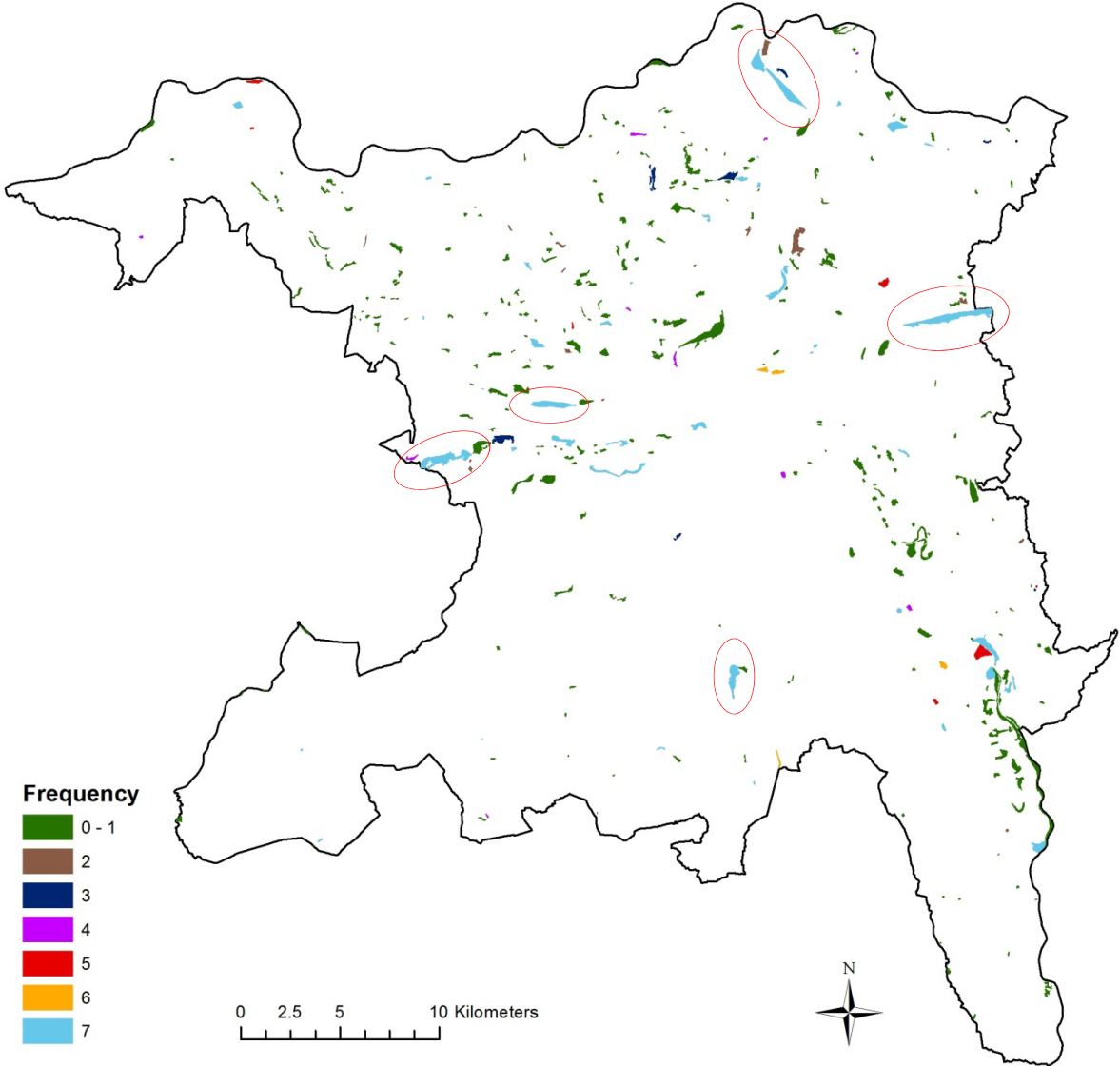


Figure 25: The selection frequency of the seven best solutions of the red list species runs was evaluated. The highest category contained 44 CCIs the lowest 268 CCIs. Some of the largest CCIs were included in every best solution of the all species runs as well as in every best solution of the red list species runs (red circles).

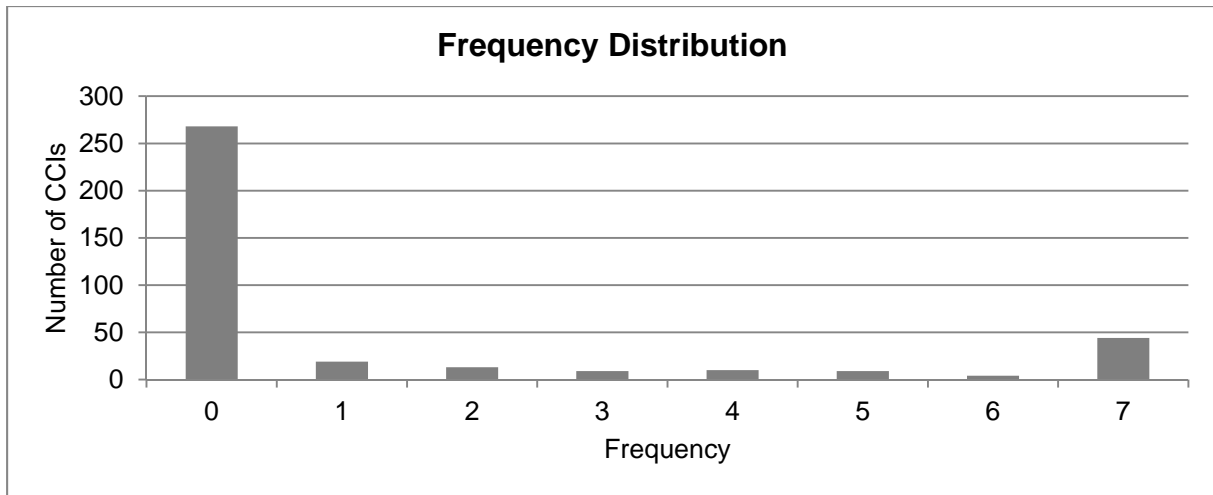


Figure 26: The the number of CCI's per selection frequency of the seven red list species runs was calculated. Here, the distribution was a more extreme version of the frequency distribution of the all species runs. The main part of the CCI's was never included; 44 CCI's were selected in every best solution.

4. Discussion

4.1 Species Data Base

When Marxan runs an optimization it is assumed that the species data are complete. But obviously in this study the data imperfectly covered the species occurrence in all the CCIs. It's about the Linnean shortfall (Whittaker, Araujo et al. 2005) that referred to our lack of information of how many, and what kind, of species there are. That means, we still have incomplete knowledge of the present biodiversity. In addition, I lacked information on the abundance of each species occurred per CCI. Thus, the results of this study should be interpreted carefully. They only illustrate which CCIs Marxan selects with the given information. Even if this analysis with Marxan gave some hints about which CCIs contribute to the protection and preservation of biodiversity, it was not meant to support decisions. This study sought to apply this systematic-conservation tool as a preliminary analysis. Its results did not intend to be implemented in practice.

In addition to the incompleteness of the species data, another species problem existed. Namely, the inventory data that I used were possibly biased in two ways. First, in this study species data were often based on field inventories of a specific species group, e.g. birds. Second, some inventories biased towards encountering rare or otherwise interesting species. In general, there is no comprehensive species inventory where all occurring species of a conservation area are recorded. Finally, there will never be a guaranty of complete species inventory even if it was sought to.

4.2 Cost Calculation

The estimations of the costs for each planning unit (here CCI) only included the current cost for relinquishment of management, compensation money and costs for maintenance. In fact, there were more costs than just these. For example, there are opportunity costs, implementation costs and even society dependent, non-monetary costs. As mentioned, the cost calculation was done only for the current cost for several reasons. It would be a challenging task to get appropriate estimations for not these additional costs that I did not consider. In addition, Marxan does not need complete cost information. It is up to the user to determine which costs Marxan uses in an optimization. If there were a specific project to plan a new or expanded conservation area network with a cost calculation for a defined time horizon, I

recommend inclusions of implementation and opportunity costs. This would result in an absolute cost for the final reserve system because it would be a more comprehensive cost calculation. In this study it was necessary to discriminate the CCIs through a meaningful cost value. Therefore, the current cost per year was considered suitable. In addition, because the cost calculation was conducted per spatial unit and then summed up for each CCI the size of the respective CCI was incorporated. Thus Marxan involved land use as well as monetary costs in the optimization. Furthermore, the applied cost calculation could be adopted as a method for future cost calculations in conservation planning in Switzerland.

4.3 Marxan Analysis Results

The outputs of Marxan indicate that there are some irreplaceable CCIs in canton Aargau. These 44 CCIs evidently contain a major part of the present biodiversity depending on the available data. Even some of the most expensive CCIs were always selected in the best solution. Hence, these CCIs seem to be essential for the reserve network and, thus, meet with the principle irreplaceability. On the contrary, there were a lot of CCIs that were never selected in a best solution. Several reasons could be figured to explain this. First, for 37 CCIs no data was available, thus they didn't contain any conservation features while they had current costs. Marxan didn't select them because it runs an optimization in consideration of benefit (conservation features) and costs (current costs). In these cases, there were only costs without benefit. Second, some CCIs contain species that could be found in another CCI at lower costs. This is more likely the fewer species a CCI contain. Hence, CCIs with few species were never selected if the species were found in another CCI at lower costs.

The analyses with Marxan indicate further that there were several species that occur in one single CCI. For this reason, the target having each species at least twice in the final reserve could not be met when the amount was set to "1". The same target was not met even when the amount was randomly set. This implies that the distribution of some species is poor in this reserve system. Obviously, the present database contributes to this conclusion.

The higher average cost per CCI of the best solutions of the red list species runs was an interesting result too. It could be casually determined as follows, if this was not an

accidental result. We assume that red list species are threatened because they make high demands on their habitat. Further, it is expensive to provide these habitats or the opportunity costs are very high. Thus, only a few adequate places for these species have remained and for which reason some species have got threatened. I recommend conducting another study to proof or disprove this thesis.

4.4 User-defined Targets

A key question in systematic conservation planning is, how many individuals of each species are required in the reserve system to ensure their persistence? This knowledge is essential when minimizing the loss of biodiversity is the objective of conservation planning. But as in this study, this knowledge is usually lacking. It would have been used to set the targets for each species in the Marxan analyses. For this reason, a set of targets was implemented in these Marxan analyses. In this way, it could be dealt with the incomplete information because the selection frequencies gave a hint of the importances of each CCI.

The arisen problem is not new for conservationists and there were several disputed attempts to formulate rules of thumb (Hanski, Moilanen et al. 1996). After Moilanen, Wilson et al. (2009) there are two approaches that account for this problem but these have limitations too. First, there are adaptable metapopulation models which operate with species-specific parameters. These models are based on the metapopulation theory (see Hanski and Gaggiotti 2004). Second, there are different types of spatial population viability analyses that integrate information on life history of a species.

There is no way to get around such modelling approaches in the field of systematic conservation planning. But it is a difficult task to decide for an appropriate method for a current problem although the methods are available. At this point, further research has to be done in Switzerland before applying systematic conservation planning in practice.

4.5 Number of Species versus Species Composition

In this study, biodiversity was only described by the number of species at a site. But as the principle of comprehensiveness (Moilanen, Wilson et al. 2009) indicates there should be consideration given to species composition and genetic diversity, to habitat types and ecological processes. These are defined as components of biodiversity as

well. While species composition and genetic diversity could be technically handled with Marxan the available information is not sufficient in this case. It is possible to run Marxan by defining the different habitat types as conservation features. This could be done in a supplementary analysis to focus on the structural diversity of conservation areas in canton Aargau. It is likely that information on ecologically important structures will remain incomplete as well. But there are several GIS-layers that would provide additional, detailed information on these additional structures. However, Marxan doesn't model ecological processes (Ardron et al. 2010). Thus, other methods have to be applied in order to address the principle of comprehensiveness in the consideration of ecological function. But it is more likely that there is a lack of appropriate data than a lack of appropriate methods.

4.6 Biodiversity Processes

One ecological process that influences genetic diversity is migration. To ensure migration of species in between conservation areas, connectivity is crucial. Since the input file for Marxan didn't offer any information of the spatial location of the CCIs, the present analysis makes no statement about the connectivity of the chosen CCIs. Thus, the effectiveness of a final reserve system concerning migration and genetic exchanges was not evaluated in this study. This problem could be solved in one of two ways. First, one could introduce a connectivity matrix to Marxan. The connectivity matrix contains numbers that account for the connectivity between planning units. If one planning unit is chosen for the solution, and the other is not, the values influence costs. Second, the whole study area of canton Aargau could be split up in a grid of regular planning units. These planning units then would share a part of their boundary with surrounding planning units. The information on the shared boundaries could be implemented in Marxan. With a boundary length modifier option one can determine how compact and thus connected the overall reserve system should be. However, these two approaches could only be applied if the database is sufficient. In this study, the data was not adequate for the connectivity matrix, nor for implementation of regular grid planning units. For these reasons the CCIs were defined as planning units even though there was this trade-off.

4.7 Dynamics

The input data for an analysis with Marxan are always snapshots in time and space. Hence the output here merely shows a near optimal solution for the moment, but not necessarily for the future. The principle of persistence is not involved directly in this analysis. But there have been studies that seek to overcome this problem. Pressey et al. (2007) dealt with the snapshot problem too. They concluded that conservation planning is not effective in providing for conservation of biodiversity processes and neither are dynamic threats considered. As an approach to include dynamics Pressey (2007) focus on modelling future conditions. This approach was implemented by Loyola et al. (2013) when they applied models to generate data about potential future ecological niches and climate. They then estimated future species occurrences based on the combination of the output of these models. Finally, they ran Marxan twice, once for the current species occurrence data and again for the future species occurrences, and subsequently compared the outputs. With this method they addressed to the problem of the static character of species occurrence data. If one projection of species occurrence was adequate is another question. Since all models make some assumptions, their output should be critically interpreted.

4.8 Transfer Gap between Science and Practice

After all the mentioned points the gap between science and practice in conservation planning in Switzerland has to be closed. Since systematic conservation planning needs a comprehensive planning process, it would take a long time until satisfactory conservation plans are on hand and could be implemented. In fact, there are concepts on how to implement systematic conservation planning in reality (see chapter 2.2 and Groves, Jensen et al. 2002). But prior to going through these steps, the idea of systematic conservation planning has to be made available for practitioners. This could be done by conducting workshops, a planning guidance or even an information center.

5. Conclusions and Future Perspectives

This study was a first attempt to run a systematic conservation-planning tool with real data on plant and animal species occurrence in a region in Switzerland. The results illustrated that Marxan could identify some CCIs that seems to be irreplaceable for the reserve network in canton Aargau. Further, the outputs of Marxan were constrained by the availability of data. Therefore, these outputs should not be used as decision support. The calculation of the current costs was founded and transparent. Due to its flexibility it could be used as a method in further studies in Switzerland. In general, this study showed a rather technical approach that needs a lot of more work to be done.

In my opinion, the concept of systematic conservation planning should be advanced in Switzerland. It is a comprehensive approach as it seeks to consider all relevant factors. Further, Marxan analyses should be used as a decision support only. However, I recommend to improve several things. First, the database of the conservation features should be more appropriate. Second, further studies should be conducted to find sufficient methods that address the problems of incomplete database, setting specific targets and including dynamics. I think, there already exist some models that could be a possibility to deal with this problems. But, the selection of such a method should be reflected for every single analysis, because each problem in conservation planning is different. Finally, there is another version of Marxan that works with spatial zonation. I suppose that Marxan with zonation has a high potential to operate with more complex problems. Thus, this software should be applied for Switzerland too.

I believe that systematic conservation planning would be best practice for Switzerland too. Therefore, it is important to close the gap between science and practice. The concept of systematic conservation planning should be made available for the Swiss conservationists in practice. This information exchange could be advanced by conducting workshops or seminars in conservation planning.

Acknowledgment

First I want to thank my supervisors Peter B. Pearman and Felix Kienast for supporting my master thesis. They organized a work place at WSL, answered to current questions and helped me clarifying some communication problems. Thanks go to Dirk Schmatz and Melanie Germann who helped in the digitalization of the collected archive data. Finally, I am thankful that Canton Aargau (Departement of Landscape and Water) provided access to the non-digitized species inventories of conservation areas of cantonal importance.

Tables

Table 1: Status value in the planning unit file	9
Table 2: Used GIS-layers of canton Aargau	10
Table 3: GIS-layer that contained field names.....	11
Table 4: Categories of red list status (source: BAFU).....	12
Table 5: Basic costs per 1ha per year	13
Table 6: Surcharge per hectare and year depending on the slope.....	13
Table 7: Surcharge or reduction per hectare and year depending on the patch size	13
Table 8: Used GIS-layers for cost calculation.....	13
Table 9: spec.dat	15
Table 10: pu.dat.....	15
Table 11: puvspr.dat.....	15
Table 12: Optimization Runs where the targets are the same for all species	16
Table 13: Red list species runs.....	17
Table 14: Summary information red list species data	18
Table 15: Similar runs grouped.....	26
Table 16: Number of missing species in the best solution.....	27
Table 17: Number of missing species in the best solutions of the red list species runs	28

Figures

Figure 1: Conservation areas of cantonal interest (green) in canton Aargau.....	4
Figure 2: The key principles for spatial prioritization problems	6
Figure 3: Simulated annealing	8
Figure 4: The number of species per CCI.....	19
Figure 5: The number of red list species per CCI	20
Figure 6: <i>Sium latifolium</i>	21
Figure 7: <i>Dactylorhiza maculata</i>	21
Figure 8: <i>Vipera aspis</i>	21
Figure 9: <i>Satyrium pruni</i>	21
Figure 10: <i>Cupido argiades</i>	21
Figure 11: <i>Leucorrhinia pectoralis</i>	21
Figure 12: <i>Leucorrhinia caudalis</i>	21
Figure 13: <i>Sympetrum pedemontanum</i>	21
Figure 14: <i>Castor fiber</i>	21
Figure 15: <i>Lanius senator</i>	22
Figure 16: <i>Vanellus vanellus</i>	22
Figure 17: The current costs per year for each CCI.....	24
Figure 18: The calculated costs.....	25
Figure 19: The calculated cost of each CCI.....	25
Figure 20: The cost and the number of selected CCIs.	26
Figure 21: The costs and number of CCIs of the best solutions.	28
Figure 22: The average cost per CCI.	29
Figure 23: The selection frequency.	30
Figure 24: The Number of CCIs.....	30
Figure 25: The selection frequency of the seven best solutions.	31
Figure 26: The the number of CCIs per selection frequency.	32

References

- ALN Amt für Landschaft und Natur des Kantons Zürich, Fachstelle Naturschutz (2013). Merkblatt Hecken
- Ardron, J.A., Possingham, H.P., and Klein, C.J. (eds). (2010). Marxan Good Practices Handbook, Version 2. Pacific Marine Analysis and Research Association, Victoria, BC, Canada. 165 pages.
- Bolliger, J., Edwards, T. C., Eggenberg, S., Ismail, S., Seidl, I., & Kienast, F. (2011). Balancing Forest-Regeneration Probabilities and Maintenance Costs in Dry Grasslands of High Conservation Priority. [Article]. *Conservation Biology*, 25(3), 567-576.
- Ferrier, S. (2002). Mapping spatial pattern in biodiversity for regional conservation planning: Where to from here? [Article; Proceedings Paper]. *Systematic Biology*, 51(2), 331-363.
- Game, E. T. and H. S. Grantham. (2008). Marxan User Manual: For Marxan version 1.8.10. University of Queensland, St. Lucia, Queensland, Australia, and Pacific Marine Analysis and Research Association, Vancouver, British Columbia, Canada.
- Groves, C. R., Jensen, D. B., Valutis, L. L., Redford, K. H., Shaffer, M. L., Scott, J. M., ... Anderson, M. G. (2002). Planning for biodiversity conservation: Putting conservation science into practice. [Article]. *Bioscience*, 52(6), 499-512.
- Hanski, I. and Gaggiotti, O. E. eds. (2004). Ecology, Genetics and Evolution of Metapopulations.
- Hanski, I., Moilanen, A., & Gyllenberg, M. (1996). Minimum viable metapopulation size. [Article]. *American Naturalist*, 147(4), 527-541.
- Ismail, S.; Schwab, F.; Tester, U.; Kienast, F.; Martinoli, D.; Seidl, I., (2009): Kosten eines gesetzeskonformen Schutzes der Biotope von nationaler Bedeutung. Technischer Bericht. Available from World Wide Web <<http://www.wsl.ch/publikationen/pdf/9625.pdf>>. Birmensdorf, Eidg. Forschungsanstalt für Wald, Schnee und Landschaft WSL; Basel, Pro Natura; Bern, Forum Biodiversität, SCNAT. 122 S.

- Keller V., Gerber A., Schmid H., Volet B., Zbinden N. (2010): Rote Liste Brutvögel. Gefährdete Arten der Schweiz, Stand 2010. Bundesamt für Umwelt, Bern, und Schweizerische Vogelwarte, Sempach. Umwelt-Vollzug Nr. 1019. 53 S.
- Landolt E., Bäumler B., Erhardt A., Hegg O., Klötzli F., Lämmli W., Nobis M., Rudmann-Maurer K., Schweingruber F.H., Theurillat J.P., Urmi E., Vust M., Wohlgemut T. (1977). Flora indicativa: Ökologische Zeigerwerte und biologische Kennzeichen zur Flora der Schweiz und der Alpen; Haupt-Verlag, 768 S.
- Loyola, R. D., Lemes, P., Nabout, J. C., Trindade, J., Sagnori, M. D., Dobrovolski, R., & Diniz, J. A. F. (2013). A straightforward conceptual approach for evaluating spatial conservation priorities under climate change. [Article]. *Biodiversity and Conservation*, 22(2), 483-495.
- Margules, C. R., & Pressey, R. L. (2000). Systematic conservation planning. [Review]. *Nature*, 405(6783), 243-253.
- McDonnell, M. D., Possingham, H. P., Ball, I. R., & Cousins, E. A. (2002). Mathematical methods for spatially cohesive reserve design. [Article]. *Environmental Modeling & Assessment*, 7(2), 107-114.
- Moilanen, A., Wilson, K. A., & Possingham, H. P. (2009). *Spatial conservation prioritization: quantitative methods and computational tools*: Oxford University Press New York.
- Monnerat C., Thorens P., Walter T., Gonseth Y. (2007): Rote Liste der Heuschrecken der Schweiz. Bundesamt für Umwelt, Bern, und Schweizer Zentrum für die Kartographie der Fauna, Neuenburg. Umwelt-Vollzug 0719: 62 S.
- Moser, D., A. Gyax, B. Bäumler, N. Wyler & R. Palese (2002): *Rote Liste der gefährdeten Farn- und Blütenpflanzen der Schweiz*. Hrsg. Bundesamt für Umwelt, Wald und Landschaft, Bern; Zentrum des Datenverbundnetzes der Schweizer Flora, Chambésy; Conservatoire et Jardin botaniques de la Ville de Genève, Chambésy. BUWAL-Reihe «Vollzug Umwelt». 118 S.

- Pearce, J. L., Kirk, D. A., Lane, C. P., Mahr, M. H., Walmsley, J., Casey, D., ... Jones, K. (2008). Prioritizing avian conservation areas for the yellowstone to Yukon Region of North America. [Article]. *Biological Conservation*, 141(4), 908-924.
- Pressey, R. L., Cabeza, M., Watts, M. E., Cowling, R. M., & Wilson, K. A. (2007). Conservation planning in a changing world. [Review]. *Trends in Ecology & Evolution*, 22(11), 583-592.
- Sarkar, S., Pressey, R. L., Faith, D. P., Margules, C. R., Fuller, T., Stoms, D. M., ... Andelman, S. (2006). Biodiversity conservation-planning tools: Present status and challenges for the future *Annual Review of Environment and Resources* (Vol. 31, pp. 123-159). Palo Alto: Annual Reviews.
- Whittaker, R. J., Araujo, M. B., Paul, J., Ladle, R. J., Watson, J. E. M., & Willis, K. J. (2005). Conservation Biogeography: assessment and prospect. [Review]. *Diversity and Distributions*, 11(1), 3-23.

Internet Sources

Bundesamt für Umwelt (BAFU):

<http://www.bafu.admin.ch/tiere/07964/08223/index.html?lang=de> (14.11.2013)

Canton Aargau:

https://www.ag.ch/de/bvu/umwelt_natur_landschaft/naturschutz/nachhaltigkeit_2/schutzgeb/sschutzgebiet_1.jsp (10.12.2013)

<https://www.ag.ch/de/bvu/wald/wald.jsp> (10.12.2013)

https://www.ag.ch/media/kanton_aargau/bvu/dokumente_2/raumentwicklung/gundlagen_6/raumb Beobachtung_1/Bericht_Bodennutzung.pdf (18.2.2014)

Fauna Europaea:

de Jong, Y.S.D.M. (ed.) (2013) Fauna Europaea version 2.6. Web Service available online at <http://www.faunaeur.org> (26.11.2013)

Info Flora:

www.infoflora.ch (08.10.2013)

Random Amounts:

www.random.org (06.02.2014)

Figures

Figure 1:

http://upload.wikimedia.org/wikipedia/commons/9/9b/Karte_Kanton_Aargau_2010.png (18.2.2014)

Figure 3:

<http://www.intechopen.com/books/simulated-annealing-single-and-multiple-objective-problems/simulated-annealing-for-fast-motion-estimation-algorithm-in-h-264-avc> (04.03.2014)

Figure 6:

<http://www.infoflora.ch/de/flora/1539-sium-latifolium.html> (11.02.2014)

Figure 7:

http://www.fourlangwebprogram.com/fourlang/nl/f_Dactylorhiza_maculata.html (11.02.2014)

Figure 8:

<http://www.artenschutz.ch/cr1.htm> (11.02.2014)

Figure 9:

<http://www.vogelwarte.ch/woodchat-shrike.html> (11.02.2014)

Figure 10:

<http://www.vogelwarte.ch/kiebitz.html> (11.02.2014)

Figure 11:

http://www.lepiforum.de/lepiwiki.pl?Satyrium_Pruni (11.02.2014)

Figure 12:

http://www.lepiforum.de/lepiwiki.pl?Cupido_Argiades (11.02.2014)

Figure 13:

<http://www.artenschutz.ch/cr3.htm> (11.02.2014)

Figure 14:

<http://www.artenschutz.ch/cr3.htm> (11.02.2014)

Figure 15:

<http://www.iucn.org/knowledge/news/?3913/Dragonflies-go-thirsty-in-the-Mediterranean> (11.02.2014)

Figure 16:

<http://www.pronatura-aargau.ch/cms/index.php?id=257> (11.02.2014)

Appendix 1

General Parameters	
VERSION	0.1
BLM	0
PROP	0
RANDSEED	-1
BESTSCORE	-1
NUMREPS	10
Annealing Parameters	
NUMITNS	1000000
STARTTEMP	-1
COOLFAC	6
NUMTEMP	10000
Cost Threshold	
COSTTRESH	0
THRESHPEN1	14
THRESHPEN2	1
Input Files	
INPUTDIR	input
SPECNAME	spec.dat
PUNAME	pu.dat
PUVSPRNAME	puvspr.dat
Save Files	
SCENNAME	output
SAVERUN	2
SAVEBEST	2
SAVESUMMARY	2
SAVESCEN	2
SAVETARGMET	2
SAVESUMSOLN	2
SAVELOG	2
SAVESNAPSTEPS	0
SAVESNAPCHANGES	0
SAVESNAPFREQUENCY	0
OUTPUTDIR	output
Program control	
RUNMODE	1
MISSLEVEL	1
ITIMPTYPE	0
HEURTYPE	-1
CLUMPTYPE	0
VERBOSITY	3
SAVESOLUTIONSMATRIX	3

Appendix 3

Plant Species	ID	Red List Status
<i>Abies alba</i>	91	LC
<i>Acer campestre</i>	1988	LC
<i>Acer opalus</i>	1989	LC
<i>Acer platanoides</i>	1986	LC
<i>Acer pseudoplatanus</i>	1985	LC
<i>Aceras anthropophorum</i>	749	VU
<i>Achillea millefolium</i>	3176	LC
<i>Achillea ptarmica</i>	3181	NT
<i>Achnatherum calamagrostis</i>	186	LC
<i>Acinos arvensis</i>	2541	LC
<i>Aconitum compactum</i>	1106	LC
<i>Aconitum lycoctonum</i>	1099	LC
<i>Acorus calamus</i>	561	VU
<i>Actaea spicata</i>	1119	LC
<i>Adenostyles alliariae</i>	3037	LC
<i>Adenostyles glabra</i>	3036	LC
<i>Adoxa moschatellina</i>	2837	LC
<i>Aegopodium podagraria</i>	2241	LC
<i>Agrimonia eupatoria</i>	1560	LC
<i>Agrostis capillaris</i>	190	LC
<i>Agrostis stolonifera</i>	191	LC
<i>Ajuga genevensis</i>	2463	LC
<i>Ajuga reptans</i>	2462	LC
<i>Alchemilla vulgaris</i>	1552	LC
<i>Alisma lanceolatum</i>	151	VU
<i>Alisma plantago-aquatica</i>	150	LC
<i>Alliaria petiolata</i>	1356	LC
<i>Allium carinatum</i>	679	NT
<i>Allium lusitanicum</i>	675	LC
<i>Allium oleraceum</i>	677	LC
<i>Allium scorodoprasum</i>	664	VU
<i>Allium sphaerocephalon</i>	667	LC
<i>Allium ursinum</i>	662	LC
<i>Allium vineale</i>	666	LC
<i>Alnus glutinosa</i>	830	LC
<i>Alnus incana</i>	831	LC
<i>Alnus viridis</i>	828	LC
<i>Alopecurus aequalis</i>	216	VU
<i>Alopecurus geniculatus</i>	215	VU
<i>Althaea hirsuta</i>	2014	EN
<i>Alyssum montanum</i>	1315	VU
<i>Amelanchier ovalis</i>	1655	LC
<i>Anacamptis pyramidalis</i>	755	VU
<i>Anagallis minima</i>	2340	EN
<i>Anemone nemorosa</i>	1137	LC
<i>Anemone ranunculoides</i>	1136	LC
<i>Angelica sylvestris</i>	2204	LC
<i>Anthemis tinctoria</i>	3166	NT
<i>Anthericum liliago</i>	631	LC
<i>Anthericum ramosum</i>	632	LC
<i>Anthoxanthum odoratum</i>	180	LC
<i>Anthriscus nitida</i>	2192	LC
<i>Anthriscus sylvestris</i>	2191	LC
<i>Anthyllis carpatica</i>	1775	LC
<i>Anthyllis vulneraria</i>	1773	LC
<i>Anthyllis vulneraria</i>	1733	LC
<i>Aquilegia atrata</i>	1114	LC
<i>Aquilegia vulgaris</i>	1113	LC
<i>Arabidopsis thaliana</i>	1405	LC
<i>Arabis hirsuta</i>	1416	LC
<i>Arabis turrata</i>	1407	LC
<i>Arctium lappa</i>	2958	LC
<i>Arenaria serpyllifolia</i>	1039	LC
<i>Arrhenatherum elatius</i>	240	LC
<i>Artemisia absinthium</i>	3222	LC
<i>Arum maculatum</i>	563	LC
<i>Aruncus dioicus</i>	1535	LC
<i>Asarum europaeum</i>	864	LC
<i>Asparagus officinalis</i>	636	LC
<i>Asperula cynanchica</i>	2775	LC
<i>Asplenium fontanum</i>	47	NT
<i>Asplenium trichomanes</i>	42	LC
<i>Asplenium viride</i>	43	LC
<i>Aster amellus</i>	3154	LC
<i>Aster bellidiastrum</i>	3162	LC
<i>Athamanta cretensis</i>	2181	LC
<i>Athyrium filix-femina</i>	30	LC
<i>Atriplex patula</i>	934	LC
<i>Atropa bella-donna</i>	2578	LC
<i>Barbarea vulgaris</i>	1389	LC
<i>Bellis perennis</i>	3163	LC
<i>Berberis vulgaris</i>	1213	LC
<i>Berula erecta</i>	2243	LC
<i>Betula pendula</i>	824	LC
<i>Bidens tripartita</i>	3106	NT
<i>Blysmus compressus</i>	434	LC
<i>Bothriochloa ischaemum</i>	161	LC
<i>Brachypodium pinnatum</i>	385	LC
<i>Brachypodium sylvaticum</i>	386	LC
<i>Briza media</i>	296	LC
<i>Bromus benekenii</i>	362	LC
<i>Bromus commutatus</i>	377	VU
<i>Bromus erectus</i>	363	NT
<i>Bromus hordeaceus</i>	370	LC
<i>Bromus inermis</i>	365	LC
<i>Bryonia dioica</i>	2884	LC
<i>Buglossoides purpureo-caerulea</i>	2454	NT
<i>Buphthalmum salicifolium</i>	3119	LC
<i>Bupleurum falcatum</i>	2151	DD
<i>Calamagrostis canescens</i>	203	VU
<i>Calamagrostis epigejos</i>	201	LC
<i>Calamagrostis varia</i>	207	LC
<i>Calamintha nepeta</i>	2540	LC
<i>Callitriche palustris</i>	1970	LC
<i>Calluna vulgaris</i>	2271	LC
<i>Caltha palustris</i>	1125	LC
<i>Calystegia sepium</i>	2403	LC
<i>Campanula glomerata</i>	2919	VU
<i>Campanula patula</i>	2936	NT
<i>Campanula persicifolia</i>	2943	LC
<i>Campanula rapunculoides</i>	2922	LC
<i>Campanula rapunculus</i>	2935	LC
<i>Campanula rotundifolia</i>	2931	LC
<i>Campanula trachelium</i>	2923	LC
<i>Capsella bursa-pastoris</i>	1276	LC
<i>Cardamine amara</i>	1371	LC
<i>Cardamine flexuosa</i>	1364	LC
<i>Cardamine heptaphylla</i>	1375	LC
<i>Cardamine hirsuta</i>	1363	LC
<i>Cardamine pentaphylla</i>	1377	LC
<i>Cardamine pratensis</i>	1369	LC
<i>Carduus defloratus</i>	2980	LC
<i>Carduus personata</i>	2985	LC
<i>Carex acuta</i>	507	LC
<i>Carex acutiformis</i>	525	LC
<i>Carex alba</i>	528	LC
<i>Carex appropinquata</i>	479	LC
<i>Carex buxbaumii</i>	500	EN
<i>Carex canescens</i>	490	LC
<i>Carex caryophyllea</i>	520	LC
<i>Carex davalliana</i>	460	LC
<i>Carex diandra</i>	478	VU
<i>Carex digitata</i>	512	LC
<i>Carex dioica</i>	461	NT
<i>Carex distans</i>	552	NT
<i>Carex disticha</i>	492	NT
<i>Carex echinata</i>	488	LC
<i>Carex elata</i>	503	LC
<i>Carex flacca</i>	522	LC
<i>Carex flava</i>	556	LC
<i>Carex hirta</i>	508	LC
<i>Carex hostiana</i>	553	LC
<i>Carex humilis</i>	511	LC
<i>Carex lasiocarpa</i>	509	NT
<i>Carex lepidocarpa</i>	557	LC
<i>Carex limosa</i>	534	NT
<i>Carex montana</i>	518	LC
<i>Carex muricata</i>	475	LC
<i>Carex nigra</i>	506	LC
<i>Carex ornithopoda</i>	513	LC
<i>Carex pallescens</i>	555	LC
<i>Carex panicea</i>	545	LC
<i>Carex paniculata</i>	480	LC
<i>Carex pendula</i>	531	LC
<i>Carex pilulifera</i>	516	LC
<i>Carex pseudocyperus</i>	530	VU
<i>Carex pulcaris</i>	458	NT
<i>Carex rostrata</i>	523	LC
<i>Carex spicata</i>	474	LC
<i>Carex sylvatica</i>	537	LC
<i>Carex tomentosa</i>	519	LC
<i>Carex umbrosa</i>	515	LC
<i>Carex vesicaria</i>	524	NT
<i>Carex viridula</i>	559	LC
<i>Carex vulpinoidea</i>	473	EN
<i>Carlina acaulis</i>	2954	LC
<i>Carlina simplex</i>	2953	LC
<i>Carlina vulgaris</i>	2951	LC
<i>Carpinus betulus</i>	822	LC
<i>Carum carvi</i>	2249	LC
<i>Centaurea grinensis</i>	3004	NT
<i>Centaurea jacea</i>	3013	NT
<i>Centaurea pannonica</i>	3014	LC
<i>Centaurea scabiosa</i>	3005	NT
<i>Centaurea stoebe</i>	3017	EN
<i>Centaureum erythraea</i>	2361	LC
<i>Centaureum pulchellum</i>	2362	VU
<i>Cephalanthera damasonium</i>	730	LC
<i>Cephalanthera longifolia</i>	729	LC
<i>Cephalanthera rubra</i>	731	LC
<i>Cerastium arvense</i>	1026	EN
<i>Cerastium fontanum</i>	1020	LC
<i>Cerastium glomeratum</i>	1018	LC
<i>Cerastium semidecandrum</i>	1012	LC
<i>Cerastium vulgare</i>	1019	LC
<i>Ceratophyllum demersum</i>	1096	VU
<i>Chaerophyllum hirsutum</i>	2187	LC
<i>Chenopodium bonus-henricus</i>	913	LC

<i>Chrysosplenium alternifolium</i>	1526	LC	<i>Dactylorhiza fuchsii</i>	778	LC	<i>Epipactis muelleri</i>	739	LC	<i>Galium aparine</i>	2814	LC	<i>Helianthemum ovatum</i>	2045	LC
<i>Cichorium intybus</i>	3232	LC	<i>Dactylorhiza incarnata</i>	775	NT	<i>Epipactis palustris</i>	734	LC	<i>Galium boreale</i>	2783	LC	<i>Helictotrichon pubescens</i>	260	LC
<i>Cicuta virosa</i>	2234	EN	<i>Dactylorhiza maculata</i>	777	CR	<i>Epipactis purpurata</i>	737	LC	<i>Galium elongatum</i>	2788	NT	<i>Helleborus foetidus</i>	1123	LC
<i>Circaea lutetiana</i>	2100	LC	<i>Dactylorhiza majalis</i>	781	LC	<i>Equisetum arvense</i>	71	LC	<i>Galium mollugo</i>	2805	LC	<i>Hemerocallis fulva</i>	634	LC
<i>Cirsium acaule</i>	2966	LC	<i>Dactylorhiza traunsteineri</i>	779	NT	<i>Equisetum fluviatile</i>	74	LC	<i>Galium odoratum</i>	2785	LC	<i>Hepatica nobilis</i>	1149	LC
<i>Cirsium arvense</i>	2961	LC	<i>Danthonia decumbens</i>	246	LC	<i>Equisetum hyemale</i>	76	LC	<i>Galium palustre</i>	2787	LC	<i>Heracleum</i>		
<i>Cirsium oleraceum</i>	2971	LC	<i>Daphne laureola</i>	2084	LC	<i>Equisetum palustre</i>	73	LC	<i>Galium pumilum</i>	2795	LC	<i>mantegazzianum</i>	2211	LC
<i>Cirsium palustre</i>	2962	LC	<i>Daphne mezereum</i>	2085	LC	<i>Equisetum sylvaticum</i>	72	LC	<i>Galium sylvaticum</i>	2812	LC	<i>Heracleum sphondylium</i>	2207	NT
<i>Cirsium spinosissimum</i>	2972	LC	<i>Daucus carota</i>	2139	LC	<i>Equisetum telmateia</i>	69	LC	<i>Galium uliginosum</i>	2790	LC	<i>Hieracium amplexicaule</i>	3347	LC
<i>Cirsium tuberosum</i>	2967	VU	<i>Deschampsia cespitosa</i>	272	LC	<i>Erigeron acer</i>	3144	DD	<i>Galium verum</i>	2803	LC	<i>Hieracium cymosum</i>	3328	NT
<i>Cirsium vulgare</i>	2960	LC	<i>Dianthus armeria</i>	990	NT	<i>Erigeron annuus</i>	3141	LC	<i>Genista germanica</i>	1682	LC	<i>Hieracium cymosum</i>	3328	NT
<i>Cladium mariscus</i>	450	NT	<i>Dianthus</i>			<i>Eriophorum angustifolium</i>	432	LC	<i>Genista tinctoria</i>	1681	LC	<i>Hieracium humile</i>	3348	LC
<i>Clematis vitalba</i>	1153	LC	<i>carthusianorum</i>	989	LC	<i>Eriophorum gracile</i>	433	EN	<i>Gentiana asclepiadea</i>	2368	LC	<i>Hieracium lactucella</i>	3333	LC
<i>Clinopodium vulgare</i>	2535	LC	<i>Dianthus superbus</i>	985	LC	<i>Eriophorum latifolium</i>	431	LC	<i>Gentiana campestris</i>	2386	LC	<i>Hieracium murorum</i>	3350	LC
<i>Colchicum autumnale</i>	641	LC	<i>Digitalis grandiflora</i>	2671	LC	<i>Erophila verna</i>	1309	LC	<i>Gentiana ciliata</i>	2383	LC	<i>Hieracium pilosella</i>	3335	LC
<i>Convallaria majalis</i>	624	LC	<i>Digitalis lutea</i>	2670	LC	<i>Erucastrum gallicum</i>	1350	NT	<i>Gentiana germanica</i>	2388	NT	<i>Hieracium piloselloides</i>	3329	LC
<i>Convolvulus arvensis</i>	2401	LC	<i>Dipsacus pilosus</i>	2862	VU	<i>Euonymus europaeus</i>	1981	LC	<i>Gentiana</i>	2367	VU	<i>Hieracium umbellatum</i>	3361	LC
<i>Cornus mas</i>	2257	LC	<i>Draba aizoides</i>	1308	LC	<i>Eupatorium cannabinum</i>	3034	LC	<i>pneumonanthe</i>	1887	LC	<i>Himantoglossum hircinum</i>	750	VU
<i>Cornus sanguinea</i>	2258	LC	<i>Draba muralis</i>	1298	VU	<i>Euphorbia amygdaloides</i>	1959	LC	<i>Geranium dissectum</i>	1887	LC	<i>Hippocrepis comosa</i>	1786	LC
<i>Coronilla coronata</i>	1784	NT	<i>Drosera anglica</i>	1444	VU	<i>Euphorbia cyparissias</i>	1960	LC	<i>Geranium malle</i>	1890	LC	<i>Hippocrepis emerus</i>	1780	LC
<i>Coronilla vaginalis</i>	1782	LC	<i>Drosera rotundifolia</i>	1443	NT	<i>Euphorbia palustris</i>	1952	VU	<i>Geranium palustre</i>	1900	NT	<i>Hippuris vulgaris</i>	2129	NT
<i>Corylus avellana</i>	821	LC	<i>Dryopteris carthusiana</i>	27	LC	<i>Euphorbia platyphyllos</i>	1953	LC	<i>Geranium pyrenaicum</i>	1891	LC	<i>Holcus lanatus</i>	238	LC
<i>Cotoneaster integerrimus</i>	1653	LC	<i>Dryopteris filix-mas</i>	22	LC	<i>Euphorbia verrucosa</i>	1957	LC	<i>Geranium robertianum</i>	1883	LC	<i>Holcus mollis</i>	239	LC
<i>Cotoneaster tomentosus</i>	1654	LC	<i>Echium vulgare</i>	2419	LC	<i>Euphrasia rostkoviana</i>	2717	LC	<i>Geranium sanguineum</i>	1895	LC	<i>Hordelymus europaeus</i>	402	LC
<i>Crataegus laevigata</i>	1656	LC	<i>Eleocharis acicularis</i>	422	VU	<i>Fagus sylvatica</i>	832	LC	<i>Geum rivale</i>	1556	LC	<i>Hottonia palustris</i>	2290	EN
<i>Crataegus monogyna</i>	1657	LC	<i>Eleocharis austriaca</i>	417	NT	<i>Festuca arundinacea</i>	339	DD	<i>Geum urbanum</i>	1557	LC	<i>Humulus lupulus</i>	849	LC
<i>Crepis biennis</i>	3318	LC	<i>Eleocharis palustris</i>	416	LC	<i>Festuca laevigata</i>	354	LC	<i>Glechoma hederacea</i>	2544	LC	<i>Hydrocharis morsus-ranae</i>	153	EN
<i>Crepis paludosa</i>	3313	LC	<i>Eleocharis quinqueflora</i>	420	LC	<i>Festuca ovina</i>	355	LC	<i>Globularia bisnagarica</i>	2755	LC	<i>Hydrocotyle vulgaris</i>	2131	VU
<i>Crepis praemorsa</i>	3310	VU	<i>Eleocharis uniglumis</i>	419	NT	<i>Festuca pratensis</i>	338	LC	<i>Globularia cordifolia</i>	2756	LC	<i>Hypericum desetangii</i>	2027	LC
<i>Crepis pyrenaica</i>	3309	LC	<i>Elodea canadensis</i>	154	LC	<i>Festuca rubra</i>	341	LC	<i>Glyceria fluitans</i>	303	LC	<i>Hypericum dubium</i>	2026	LC
<i>Crepis taraxacifolia</i>	3322	LC	<i>Elymus repens</i>	390	LC	<i>Filipendula ulmaria</i>	1563	LC	<i>Glyceria maxima</i>	301	VU	<i>Hypericum maculatum</i>	2025	LC
<i>Crepis vesicaria</i>	3323	LC	<i>Epilobium angustifolium</i>	2105	LC	<i>Filipendula vulgaris</i>	1564	VU	<i>Glyceria notata</i>	304	LC	<i>Hypericum montanum</i>	2031	LC
<i>Crocus albiflorus</i>	701	LC	<i>Epilobium collinum</i>	2110	LC	<i>Fragaria vesca</i>	1568	LC	<i>Goodyera repens</i>	741	LC	<i>Hypericum perforatum</i>	2023	LC
<i>Cucubalus baccifer</i>	975	VU	<i>Epilobium dodonaei</i>	2106	LC	<i>Frangula alnus</i>	2001	LC	<i>Groenlandia densa</i>	124	NT	<i>Hypericum perfoliatum</i>	2023	LC
<i>Cuscuta epithymum</i>	2406	LC	<i>Epilobium dodonaei</i>	2106	LC	<i>Fraxinus excelsior</i>	2354	LC	<i>Gymnadenia conopsea</i>	756	LC	<i>Hypochaeris radicata</i>	3256	LC
<i>Cynosurus cristatus</i>	236	LC	<i>Epilobium hirsutum</i>	2109	LC	<i>Galanthus nivalis</i>	696	NT	<i>Gymnadenia odoratissima</i>	757	LC	<i>Ilex aquifolium</i>	1979	LC
<i>Cyperus flavescens</i>	405	VU	<i>Epilobium montanum</i>	2111	LC	<i>Galeopsis angustifolia</i>	2496	NT	<i>Gymnocarpium dryopteris</i>	18	LC	<i>Impatiens glandulifera</i>	1993	LC
<i>Cyperus fuscus</i>	406	VU	<i>Epilobium palustre</i>	2114	LC	<i>Galeopsis bifida</i>	2502	VU	<i>Hedera helix</i>	2130	LC	<i>Impatiens noli-tangere</i>	1991	LC
<i>Cypripedium calceolus</i>	717	VU	<i>Epilobium parviflorum</i>	2108	LC	<i>Galeopsis tetrahit</i>	2501	LC	<i>Helianthemum grandiflorum</i>	2046	LC	<i>Impatiens parviflora</i>	1992	LC
<i>Cystopteris fragilis</i>	35	LC	<i>Epilobium tetragonum</i>	2122	NT	<i>Galium album</i>	2806	LC	<i>Helianthemum nummularium</i>	2044	LC	<i>Inula conyzae</i>	3124	LC
<i>Dactylis glomerata</i>	294	LC	<i>Epipactis atrorubens</i>	735	LC	<i>Galium anisophyllum</i>	2797	LC				<i>Inula graveolens</i>	3122	DD
			<i>Epipactis helleborine</i>	738	LC							<i>Inula salicina</i>	3131	NT
			<i>Epipactis microphylla</i>	736	NT							<i>Iris pseudacorus</i>	706	LC

<i>Iris sibirica</i>	707	VU	<i>Leucosium vernum</i>	697	LC	<i>Melica uniflora</i>	235	LC	<i>Orchis ustulata</i>	764	NT	<i>Platanthera bifolia</i>	753	LC	
<i>Isoplepis setacea</i>	443	VU	<i>Ligustrum vulgare</i>	2351	LC	<i>Melilotus albus</i>	1740	LC	<i>Origanum majorana</i>	2550	DD	<i>Platanthera chlorantha</i>	754	LC	
<i>Juglans regia</i>	820	LC	<i>Lilium bulbiferum</i>	652	NT	<i>Melilotus officinalis</i>	1741	LC	<i>Origanum vulgare</i>	2549	LC	<i>Poa angustifolia</i>	325	LC	
<i>Juncus acutiflorus</i>	600	NT	<i>Lilium martagon</i>	650	LC	<i>Melittis melissophyllum</i>	2547	LC	<i>Ornithogalum umbellatum</i>	656	LC	<i>Poa annua</i>	307	LC	
<i>Juncus articulatus</i>	599	LC	<i>Limodorum abortivum</i>	719	NT	<i>Mentha aquatica</i>	2563	LC	<i>Orobanchaceae</i>	2733	LC	<i>Poa bulbosa</i>	311	LC	
<i>Juncus bufonius</i>	581	LC	<i>Linaria vulgaris</i>	2650	LC	<i>Mentha arvensis</i>	2562	LC	<i>Orobanchaceae caryophyllacea</i>	2733	LC	<i>Poa palustris</i>	320	LC	
<i>Juncus conglomeratus</i>	578	LC	<i>Linum catharticum</i>	1914	LC	<i>Mentha longifolia</i>	2565	LC	<i>Orobanchaceae elatior</i>	2738	EN	<i>Poa pratensis</i>	324	LC	
<i>Juncus effusus</i>	579	LC	<i>Linum tenuifolium</i>	1916	LC	<i>Menyanthes trifoliata</i>	2357	LC	<i>Orobanchaceae minor</i>	2737	LC	<i>Poa trivialis</i>	309	VU	
<i>Juncus fuscoater</i>	598	LC	<i>Liparis loeselii</i>	744	VU	<i>Mercurialis perennis</i>	1940	LC	<i>Orobanchaceae reticulata</i>	2732	LC	<i>Polygala amara</i>	1351	DD	
<i>Juncus inflexus</i>	577	LC	<i>Listera ovata</i>	747	LC	<i>Milium effusum</i>	209	LC	<i>Orobanchaceae teucrii</i>	2743	LC	<i>Polygala amarella</i>	1932	LC	
<i>Juncus subnodulosus</i>	597	LC	<i>Lithospermum officinale</i>	2455	NT	<i>Moehringia muscosa</i>	1045	LC	<i>Orthilia secunda</i>	2261	LC	<i>Polygala comosa</i>	1936	LC	
<i>Juniperus communis</i>	101	LC	<i>Lolium multiflorum</i>	381	LC	<i>Moehringia trinervia</i>	1041	LC	<i>Oxalis acetosella</i>	1909	LC	<i>Polygala vulgaris</i>	1934	NT	
<i>Kerneria saxatilis</i>	1297	LC	<i>Lolium perenne</i>	380	LC	<i>Molinia arundinacea</i>	282	LC	<i>Oxalis stricta</i>	1912	NA	<i>Polygonatum multiflorum</i>	626	LC	
<i>Knautia arvensis</i>	2868	LC	<i>Lonicera alpigena</i>	2827	LC	<i>Molinia caerulea</i>	281	LC	<i>Papaver rhoeas</i>	1223	LC	<i>Polygonatum odoratum</i>	627	LC	
<i>Knautia dipsacifolia</i>	2867	LC	<i>Lonicera nigra</i>	2830	LC	<i>Monotropa hypopitys</i>	2267	LC	<i>Paris quadrifolia</i>	622	LC	<i>Polygonatum verticillatum</i>	625	LC	
<i>Koeleria macrantha</i>	288	LC	<i>Lonicera periclymenum</i>	2832	NT	<i>Muscari comosum</i>	689	LC	<i>Parnassia palustris</i>	1525	LC	<i>Polygonum amphibium</i>	906	NT	
<i>Koeleria pyramidata</i>	286	LC	<i>Lonicera xylosteum</i>	2829	LC	<i>Muscari racemosum</i>	687	NT	<i>Pastinaca sativa</i>	2212	LC	<i>Polygonum aviculare</i>	899	LC	
<i>Lactuca perennis</i>	3283	LC	<i>Lotus corniculatus</i>	1763	LC	<i>Mycelis muralis</i>	3290	LC	<i>Pedicularis palustris</i>	2676	LC	<i>Polygonum bistorta</i>	897	LC	
<i>Lactuca serriola</i>	3288	LC	<i>Lotus maritimus</i>	1767	LC	<i>Myosotis arvensis</i>	2451	LC	<i>Pedicularis sylvatica</i>	2677	VU	<i>Polystichum aculeatum</i>	14	LC	
<i>Lamium galeobdolon</i>	2508	LC	<i>Lotus pedunculatus</i>	1762	LC	<i>Myosotis scorpioides</i>	2441	LC	<i>Petasites albus</i>	3041	LC	<i>Populus alba</i>	784	LC	
<i>Lamium maculatum</i>	2512	LC	<i>Lunaria rediviva</i>	1290	LC	<i>Myosotis sylvatica</i>	2445	LC	<i>Petrorhagia prolifera</i>	984	LC	<i>Populus nigra</i>	782	LC	
<i>Lamium purpureum</i>	2513	LC	<i>Luzula campestris</i>	613	LC	<i>Myosoton aquaticum</i>	999	LC	<i>Peucedanum cervaria</i>	2218	LC	<i>Populus tremula</i>	783	LC	
<i>Larix decidua</i>	100	LC	<i>Luzula multiflora</i>	614	LC	<i>Myriophyllum spicatum</i>	2126	NT	<i>Peucedanum oreoselinum</i>	2217	LC	<i>Potamogeton crispus</i>	123	LC	
<i>Laserpitium latifolium</i>	2162	LC	<i>Luzula pilosa</i>	603	LC	<i>Nasturtium officinale</i>	1378	LC	<i>Peucedanum palustre</i>	2225	NT	<i>Potamogeton gramineus</i>	126	EN	
<i>Laserpitium siler</i>	2160	LC	<i>Luzula sylvatica</i>	609	LC	<i>Neottia nidus-avis</i>	721	LC	<i>Phalaris arundinacea</i>	178	LC	<i>Potamogeton natans</i>	116	LC	
<i>Lathraea squamaria</i>	2725	LC	<i>Lycopodium annotinum</i>	80	LC	<i>Nuphar lutea</i>	1092	LC	<i>Phleum bertolonii</i>	223	LC	<i>Potamogeton perfoliatus</i>	121	LC	
<i>Lathyrus linifolius</i>	1868	LC	<i>Lycopus europaeus</i>	2558	EN	<i>Nymphaea alba</i>	1090	NT	<i>Phleum pratense</i>	222	LC	<i>Potentilla anserina</i>	1580	LC	
<i>Lathyrus niger</i>	1874	LC	<i>Lysimachia nemorum</i>	2331	LC	<i>Nymphoides peltata</i>	2358	VU	<i>Phragmites australis</i>	242	LC	<i>Potentilla erecta</i>	1606	LC	
<i>Lathyrus pratensis</i>	1861	LC	<i>Lysimachia nummularia</i>	2332	LC	<i>Onobrychis viciifolia</i>	1817	LC	<i>Phyllitis scolopendrium</i>	56	LC	<i>Potentilla neumanniana</i>	1601	LC	
<i>Lathyrus sylvestris</i>	1865	LC	<i>Lysimachia thyrsoiflora</i>	2335	VU	<i>Ononis repens</i>	1704	LC	<i>Phyteuma orbiculare</i>	2899	LC	<i>Potentilla palustris</i>	1566	LC	
<i>Lathyrus vernus</i>	1872	NT	<i>Lysimachia vulgaris</i>	2334	LC	<i>Ononis rotundifolia</i>	1700	LC	<i>Phyteuma spicatum</i>	2906	LC	<i>Potentilla pratensis</i>	1587	LC	
<i>Leersia oryzoides</i>	230	EN	<i>Lythrum salicaria</i>	2091	LC	<i>Ononis spinosa</i>	1703	NT	<i>Picea abies</i>	92	LC	<i>Potentilla recta</i>	1587	LC	
<i>Legousia speculum-veneris</i>	2909	VU	<i>Malus sylvestris</i>	1666	NT	<i>Onopordum acanthium</i>	2987	VU	<i>Picris hieracioides</i>	3269	LC	<i>Potentilla reptans</i>	1605	LC	
<i>Lemna minor</i>	569	LC	<i>Malva moschata</i>	2008	LC	<i>Ophioglossum vulgatum</i>	59	VU	<i>Pimpinella major</i>	2245	LC	<i>Potentilla sterilis</i>	1577	LC	
<i>Lemna trisulca</i>	567	NT	<i>Medicago falcata</i>	1757	LC	<i>Ophrys apifera</i>	726	EN	<i>Pimpinella saxifraga</i>	2246	LC	<i>Prenanthes purpurea</i>	3282	LC	
<i>Leontodon autumnalis</i>	3265	LC	<i>Medicago lupulina</i>	1749	LC	<i>Ophrys holosericea</i>	724	EN	<i>Pinguicula vulgaris</i>	2747	LC	<i>Primula auricula</i>	2304	LC	
<i>Leontodon hispidus</i>	3263	LC	<i>Medicago minima</i>	1752	LC	<i>Ophrys insectifera</i>	727	NT	<i>Pinus sylvestris</i>	96	LC	<i>Primula elatior</i>	2298	LC	
<i>Leucanthemum adustum</i>	3203	LC	<i>Medicago sativa</i>	1756	LC	<i>Ophrys sphegodes</i>	728	EN	<i>Plantago atrata</i>	2764	LC	<i>Primula farinosa</i>	2302	LC	
<i>Leucanthemum ircutianum</i>	3199	LC	<i>Melampyrum arvense</i>	2701	VU	<i>Orchis mascula</i>	773	LC	<i>Plantago lanceolata</i>	2761	LC	<i>Primula veris</i>	2300	LC	
<i>Leucanthemum ircutianum</i>	3197	LC	<i>Melampyrum pratense</i>	2704	LC	<i>Orchis militaris</i>	767	NT	<i>Plantago major</i>	2758	LC	<i>Primula veris ssp columnae</i>	2301	LC	
			<i>Melampyrum pratense</i>	2705	LC	<i>Orchis morio</i>	762	NT	<i>Plantago media</i>	2760	LC				
			<i>Melica ciliata</i>	232	LC	<i>Orchis pallens</i>	769	NT							
			<i>Melica nutans</i>	234	LC	<i>Orchis purpurea</i>	766	VU							

<i>Prunella grandiflora</i>	2495	LC
<i>Prunella laciniata</i>	2493	EN
<i>Prunella vulgaris</i>	2494	LC
<i>Prunus avium</i>	1648	LC
<i>Prunus domestica</i>	1647	LC
<i>Prunus padus</i>	1637	NT
<i>Prunus spinosa</i>	1644	LC
<i>Pteridium aquilinum</i>	8	LC
<i>Pulicaria dysenterica</i>	3134	LC
<i>Pulmonaria officinalis</i>	2434	LC
<i>Pulsatilla vulgaris</i>	1146	EN
<i>Pyrola rotundifolia</i>	2264	LC
<i>Pyrus pyraster</i>	1665	LC
<i>Quercus petraea</i>	835	LC
<i>Quercus pubescens</i>	836	LC
<i>Quercus robur</i>	834	LC
<i>Ranunculus aconitifolius</i>	1171	LC
<i>Ranunculus acris</i>	1207	LC
<i>Ranunculus acris</i> ssp <i>friesianus</i>	1206	LC
<i>Ranunculus auricomus</i>	1185	LC
<i>Ranunculus bulbosus</i>	1192	LC
<i>Ranunculus flammula</i>	1178	NT
<i>Ranunculus fluitans</i>	1161	NT
<i>Ranunculus lingua</i>	1177	VU
<i>Ranunculus repens</i>	1193	LC
<i>Ranunculus trichophyllus</i>	1162	NT
<i>Ranunculus tuberosus</i>	1195	LC
<i>Raphanus raphanistrum</i>	1335	LC
<i>Reseda lutea</i>	1439	LC
<i>Reseda luteola</i>	1440	VU
<i>Reynoutria japonica</i>	895	LC
<i>Rhamnus alpina</i>	1998	LC
<i>Rhamnus cathartica</i>	1996	LC
<i>Rhinanthus alectorolophus</i>	2694	LC
<i>Rhinanthus angustifolius</i>	2695	VU
<i>Rhinanthus glacialis</i>	2697	LC
<i>Rhinanthus minor</i>	2699	LC
<i>Rhynchospora alba</i>	451	NT
<i>Ribes rubrum</i>	1531	LC
<i>Ribes uva-crispa</i>	1528	LC

<i>Robinia pseudoacacia</i>	1769	LC
<i>Rorippa amphibia</i>	1385	VU
<i>Rorippa islandica</i>	1381	NT
<i>Rosa arvensis</i>	1614	LC
<i>Rosa canina</i>	1630	LC
<i>Rosa glauca</i>	1618	LC
<i>Rosa jundzillii</i>	1622	NT
<i>Rosa pendulina</i>	1616	LC
<i>Rosa spinosissima</i>	1615	LC
<i>Rosa tomentella</i>	1632	VU
<i>Rosa tomentosa</i>	1624	LC
<i>Rubus caesius</i>	1610	LC
<i>Rubus fruticosus</i>	1611	LC
<i>Rubus idaeus</i>	1609	LC
<i>Rubus saxatilis</i>	1608	LC
<i>Rumex acetosa</i>	876	LC
<i>Rumex acetosella</i>	871	LC
<i>Rumex hydrolapathum</i>	883	EN
<i>Rumex obtusifolius</i>	888	LC
<i>Sagittaria latifolia</i>	143	VU
<i>Sagittaria sagittifolia</i>	142	EN
<i>Salix alba</i>	793	LC
<i>Salix appendiculata</i>	814	LC
<i>Salix aurita</i>	817	LC
<i>Salix caprea</i>	815	LC
<i>Salix cinerea</i>	818	LC
<i>Salix elaeagnos</i>	791	LC
<i>Salix myrsinifolia</i>	800	LC
<i>Salix purpurea</i>	789	LC
<i>Salix repens</i>	796	NT
<i>Salix triandra</i>	805	LC
<i>Salix viminalis</i>	792	LC
<i>Salvia glutinosa</i>	2473	LC
<i>Salvia pratensis</i>	2477	LC
<i>Sambucus nigra</i>	2821	LC
<i>Sambucus racemosa</i>	2822	LC
<i>Sanguisorba minor</i>	1538	NT
<i>Sanguisorba officinalis</i>	1536	LC
<i>Sanicula europaea</i>	2138	LC
<i>Saponaria officinalis</i>	979	LC
<i>Saxifraga paniculata</i>	1487	LC
<i>Saxifraga tridactylites</i>	1522	LC
<i>Scabiosa columbaria</i>	2879	LC
<i>Schoenoplectus</i>	448	LC

<i>lacustris</i>		
<i>Schoenoplectus tabernaemontani</i>	449	VU
<i>Schoenus ferrugineus</i>	414	NT
<i>Schoenus nigricans</i>	413	NT
<i>Scilla bifolia</i>	653	LC
<i>Scirpus sylvaticus</i>	436	LC
<i>Scrophularia canina</i>	2666	LC
<i>Scrophularia nodosa</i>	2663	LC
<i>Scutellaria galericulata</i>	2482	LC
<i>Securigera varia</i>	1785	LC
<i>Sedum acre</i>	1476	LC
<i>Sedum album</i>	1474	LC
<i>Sedum rubens</i>	1478	VU
<i>Sedum sexangulare</i>	1477	LC
<i>Sedum telephium</i>	1460	LC
<i>Selinum carvifolia</i>	2202	VU
<i>Senecio aquaticus</i>	3097	NT
<i>Senecio erucifolius</i>	3094	LC
<i>Senecio jacobaea</i>	3095	LC
<i>Senecio paludosus</i>	3081	NT
<i>Serratula tinctoria</i>	2994	NT
<i>Seseli libanotis</i>	2175	LC
<i>Sesleria caerulea</i>	251	LC
<i>Silaum silaus</i>	2184	NT
<i>Silene dioica</i>	954	LC
<i>Silene flos-cuculi</i>	953	LC
<i>Silene nutans</i>	959	NT
<i>Silene pratensis</i>	955	LC
<i>Silene vulgaris</i>	965	NT
<i>Sinapis arvensis</i>	1336	LC
<i>Sium latifolium</i>	2244	CR
<i>Solanum dulcamara</i>	2569	LC
<i>Solidago canadensis</i>	3136	LC
<i>Solidago virgaurea</i>	3138	LC
<i>Sonchus asper</i>	3296	LC
<i>Sonchus oleraceus</i>	3295	LC
<i>Sorbus aria</i>	1663	LC
<i>Sorbus aucuparia</i>	1659	LC
<i>Sorbus mougeotii</i>	1664	LC
<i>Sorbus torminalis</i>	1661	LC
<i>Sparganium emersum</i>	113	VU
<i>Sparganium erectum</i>	110	EN
<i>Spiranthes aestivalis</i>	733	VU

<i>Spiranthes spiralis</i>	732	NT
<i>Stachys annua</i>	2517	VU
<i>Stachys officinalis</i>	2527	VU
<i>Stachys palustris</i>	2520	NT
<i>Stachys recta</i>	2518	VU
<i>Stachys sylvatica</i>	2521	LC
<i>Stellaria graminea</i>	1007	LC
<i>Stellaria media</i>	1002	LC
<i>Stratiotes aloides</i>	159	VU
<i>Succisa pratensis</i>	2864	LC
<i>Symphytum officinale</i>	2432	LC
<i>Tamux communis</i>	690	LC
<i>Tanacetum corymbosum</i>	3191	NT
<i>Tanacetum vulgare</i>	3193	LC
<i>Taraxacum officinale</i>	3272	LC
<i>Taraxacum palustre</i>	3271	LC
<i>Taxus baccata</i>	90	LC
<i>Teucrium chamaedrys</i>	2468	LC
<i>Teucrium montanum</i>	2466	LC
<i>Teucrium scordium</i>	2469	EN
<i>Teucrium scorodonia</i>	2471	LC
<i>Thalictrum aquilegifolium</i>	1126	LC
<i>Thalictrum flavum</i>	1133	VU
<i>Thalictrum minus</i>	1128	NT
<i>Thelypteris palustris</i>	21	VU
<i>Thesium alpinum</i>	860	LC
<i>Thesium bavarum</i>	858	NT
<i>Thesium linophyllum</i>	856	VU
<i>Thesium pyrenaicum</i>	861	LC
<i>Thlaspi montanum</i>	1269	LC
<i>Thlaspi perfoliatum</i>	1268	LC
<i>Thymus oenipontanus</i>	2555	LC
<i>Thymus praecox</i> ssp. <i>polytrichus</i>	2554	LC
<i>Thymus pulegioides</i>	2556	LC
<i>Thymus pulegioides</i> ssp. <i>carniolicus</i>	2557	LC
<i>Tilia cordata</i>	2005	LC
<i>Tilia platyphyllos</i>	2006	LC
<i>Tofieldia calyculata</i>	618	LC
<i>Torilis japonica</i>	2145	LC
<i>Tragopogon pratensis</i>	3240	LC

<i>Tragopogon pratensis</i> ssp. <i>orientalis</i>	3239	LC
<i>Trichophorum cespitosum</i>	426	LC
<i>Trifolium aureum</i>	1708	NT
<i>Trifolium campestre</i>	1710	LC
<i>Trifolium dubium</i>	1711	LC
<i>Trifolium fragiferum</i>	1714	VU
<i>Trifolium medium</i>	1735	LC
<i>Trifolium montanum</i>	1718	LC
<i>Trifolium pratense</i>	1737	LC
<i>Trifolium repens</i>	1723	VU
<i>Trifolium rubens</i>	1730	NT
<i>Trifolium thalii</i>	1724	LC
<i>Triglochin palustris</i>	141	LC
<i>Trisetum flavescens</i>	277	LC
<i>Trollius europaeus</i>	1120	LC
<i>Tussilago farfara</i>	3043	LC
<i>Typha angustifolia</i>	108	NT
<i>Typha latifolia</i>	106	LC
<i>Typha minima</i>	109	EN
<i>Typha shuttleworthii</i>	107	VU
<i>Ulmus glabra</i>	841	LC
<i>Ulmus minor</i>	840	NT
<i>Urtica dioica</i>	851	LC
<i>Utricularia minor</i>	2751	VU
<i>Vaccinium myrtillus</i>	2281	LC
<i>Vaccinium oxycoccos</i>	2277	NT
<i>Vaccinium uliginosum</i>	2283	LC
<i>Vaccinium vitis-idaea</i>	2280	LC
<i>Valeriana dioica</i>	2846	LC
<i>Valeriana montana</i>	2844	LC
<i>Valeriana officinalis</i>	2848	LC
<i>Valeriana repens</i>	2852	LC
<i>Valeriana wallrothii</i>	2850	VU
<i>Valerianella locusta</i>	2855	LC
<i>Verbascum lychnitis</i>	2595	LC
<i>Verbascum nigrum</i>	2597	LC
<i>Verbascum thapsus</i>	2589	LC
<i>Verberna officinalis</i>	2461	LC
<i>Veronica anagallis-aquatica</i>	2603	LC
<i>Veronica arvensis</i>	2624	LC
<i>Veronica beccabunga</i>	2602	LC

<i>Veronica chamaedrys</i>	2614	LC
<i>Veronica filiformis</i>	2634	LC
<i>Veronica persica</i>	2633	LC
<i>Veronica polita</i>	2630	LC
<i>Veronica scutellata</i>	2611	VU
<i>Veronica serpyllifolia</i>	2621	LC
<i>Veronica spicata</i>	2638	LC

<i>Veronica teucrium</i>	2606	LC
<i>Viburnum lantana</i>	2824	LC
<i>Viburnum opulus</i>	2825	LC
<i>Vicia cracca</i>	1825	NT
<i>Vicia cracca</i> ssp. <i>tenuifolia</i>	1827	NT
<i>Vicia dumetorum</i>	1832	NT

<i>Vicia hirsuta</i>	1821	LC
<i>Vicia sativa</i>	1840	LC
<i>Vicia sativa</i> ssp. <i>nigra</i>	1844	LC
<i>Vicia sepium</i>	1835	LC
<i>Vicia sylvatica</i>	1830	LC
<i>Vicia tetrasperma</i>	1822	NT
<i>Vicia villosa</i>	1829	VU

<i>Vinca minor</i>	2397	LC
<i>Vincetoxicum</i> <i>hirundinaria</i>	2399	LC
<i>Viola alba</i>	2069	VU
<i>Viola canina</i>	2076	NT
<i>Viola hirta</i>	2065	LC
<i>Viola palustris</i>	2063	LC

<i>Viola reichenbachiana</i>	2074	LC
<i>Viola tricolor</i>	2053	LC
<i>Zannichellia palustris</i>	135	VU

Animal Species	ID	Red List Status
<i>Accipiter nisus</i>	96716	LC
<i>Acrocephalus arundinaceus</i>	97319	NT
<i>Acrocephalus palustris</i>	97317	LC
<i>Acrocephalus schoenobaenus</i>	97314	NE
<i>Acrocephalus scirpaceus</i>	97318	LC
<i>Acroloxus lacustris</i>	421525	LC
<i>Acronicta aceris</i>	449422	NA
<i>Acronicta psi</i>	449420	NA
<i>Actitis hypoleucos</i>	96856	EN
<i>Aculepeira ceropegia</i>	348377	NA
<i>Adscita statices</i>	440602	NA
<i>Aegithalos caudatus</i>	97278	LC
<i>Aeshna affinis</i>	214299	NE
<i>Aeshna cyanea</i>	214303	LC
<i>Aeshna isosceles</i>	214305	LC
<i>Aeshna mixta</i>	214308	LC
<i>Agapanthia violacea</i>	114168	NA
<i>Aglais io</i>	441678	NA
<i>Aglais urticae</i>	441679	NA
<i>Aglia tau</i>	443627	NA
<i>Agrius convolvuli</i>	443845	NA
<i>Agrotis ipsilon</i>	448502	NA
<i>Alauda arvensis</i>	97404	NT
<i>Alcedo atthis</i>	97082	VU
<i>Alsophila aceraria</i>	446361	NA
<i>Alytes obstetricans</i>	177873	EN
<i>Ampedus sinuatus</i>	235549	NA
<i>Anas crecca</i>	96500	VU
<i>Anas platyrhynchos</i>	96502	LC
<i>Anas strepera</i>	96498	EN
<i>Anax imperator</i>	214314	LC
<i>Anax parthenope</i>	214316	LC
<i>Anguis fragilis</i>	214587	LC
<i>Anisus leucostoma</i>	430524	LC
<i>Anthocharis cardamines</i>	440867	NA
<i>Anthus trivialis</i>	97417	LC
<i>Apatura iris</i>	441662	VU

<i>Aplocera plagiata</i>	444530	NA
<i>Aporia crataegi</i>	440897	VU
<i>Araschnia levana</i>	441672	NA
<i>Ardea cinerea</i>	96658	LC
<i>Argiope bruennichi</i>	348443	NE
<i>Argynnis paphia</i>	441744	NA
<i>Aricia agestis</i>	441006	VU
<i>Asio otus</i>	97051	NT
<i>Auchmis detera</i>	447366	NA
<i>Autographa gamma</i>	449605	NA
<i>Aythya ferina</i>	96513	EN
<i>Aythya fuligula</i>	96519	VU
<i>Boloria dia</i>	441716	EN
<i>Boloria euphrosyne</i>	441700	NA
<i>Bombina variegata</i>	177877	EN
<i>Bombus sylvarum</i>	231919	NA
<i>Brachytron pratense</i>	214318	LC
<i>Brenthis ino</i>	441735	VU
<i>Bufo bufo</i>	177878	VU
<i>Bufo calamita</i>	177879	EN
<i>Buteo buteo</i>	96722	LC
<i>Calliptamus italicus</i>	402751	VU
<i>Callitarea pudibunda</i>	447103	NA
<i>Callophrys rubi</i>	441177	VU
<i>Calopteryx splendens</i>	214240	LC
<i>Carduelis cannabina</i>	97472	NT
<i>Carduelis chloris</i>	97465	LC
<i>Carduelis spinus</i>	97471	LC
<i>Carterocephalus palaemon</i>	440735	NA
<i>Castor fiber</i>	305558	CR
<i>Catocala fulminea</i>	446652	NA
<i>Catocala promissa</i>	446678	NA
<i>Cerambyx scopolii</i>	114746	NA
<i>Cercion lindenii</i>	214258	NT
<i>Certhia brachydactyla</i>	97261	LC
<i>Chalicedoma parietina</i>	231979	VU
<i>Charadrius dubius</i>	96870	EN
<i>Chorthippus albomarginatus</i>	402549	LC
<i>Chorthippus mollis</i>	402587	NT
<i>Chrysochraon dispar</i>	402547	NT
<i>Cicadetta montana</i>	239538	NA
<i>Cicindela germanica</i>	387305	EN

<i>Ciconia ciconia</i>	96674	VU
<i>Cinclus cinclus</i>	97156	LC
<i>Coccothraustes coccothraustes</i>	97498	LC
<i>Coenagrion puella</i>	214272	LC
<i>Coenonympha arcania</i>	441264	NT
<i>Coenonympha pamphilus</i>	441285	NA
<i>Coenonympha tullia</i>	441253	EN
<i>Colias croceus</i>	440816	NA
<i>Colias hyale</i>	440826	NA
<i>Columba oenas</i>	97003	LC
<i>Columba palumbus</i>	97005	LC
<i>Conocephalus fuscus</i>	403179	VU
<i>Cordulegaster bidentata</i>	214348	NT
<i>Cordulegaster boltonii</i>	214349	LC
<i>Cordulia aenea</i>	214357	LC
<i>Coronella austriaca</i>	214732	VU
<i>Corvus corax</i>	97146	LC
<i>Corvus corone</i>	97143	LC
<i>Crocothemis erythraea</i>	214375	LC
<i>Cucullia umbratica</i>	449318	NA
<i>Cuculus canorus</i>	97028	NT
<i>Cupido argiades</i>	441120	CR
<i>Cupido minimus</i>	441111	VU
<i>Cyaniris semiargus</i>	440997	NA
<i>Decticus verrucivorus</i>	403450	NT
<i>Deilephila porcellus</i>	443741	NA
<i>Dendrocopos major</i>	97099	LC
<i>Dendrocopos medius</i>	97101	NT
<i>Dendrocopos minor</i>	97103	LC
<i>Diachrysa chrysis</i>	449550	NA
<i>Dolomedes fimbriatus</i>	352481	NA
<i>Dryocopus martius</i>	97095	LC
<i>Ectropis crepuscularia</i>	445706	NA
<i>Emberiza cirulus</i>	97524	NT
<i>Emberiza citrinella</i>	97523	LC
<i>Emberiza schoeniclus</i>	97536	VU
<i>Erebia aethiops</i>	441347	VU
<i>Erebia ligea</i>	441321	NA
<i>Erinaceus europaeus</i>	305459	LC
<i>Eriogaster lanestris</i>	443530	NA
<i>Erithacus rubecula</i>	97170	LC

<i>Erynnis tages</i>	440801	NA
<i>Erythromma najas</i>	214278	LC
<i>Erythromma viridulum</i>	214279	LC
<i>Euclidia glyphica</i>	446607	NA
<i>Euconulus alderi</i>	425745	NT
<i>Euthystira brachyptera</i>	402521	LC
<i>Euxoa nigrofusca</i>	448566	NA
<i>Falco subbuteo</i>	96742	NT
<i>Falco tinnunculus</i>	96738	NT
<i>Favonius quercus</i>	441154	NA
<i>Ficedula hypoleuca</i>	97165	LC
<i>Formica polyctena</i>	81321	NT
<i>Formica rufa</i>	81325	NT
<i>Fringilla coelebs</i>	97456	LC
<i>Fringilla montifringilla</i>	97459	NA
<i>Fulica atra</i>	96786	LC
<i>Gallinula chloropus</i>	96779	LC
<i>Garrulus glandarius</i>	97127	LC
<i>Gerris lacustris</i>	450573	NA
<i>Glis glis</i>	305715	LC
<i>Gomphus vulgatissimus</i>	214334	NT
<i>Gonepteryx rhamni</i>	440831	NA
<i>Gryllus campestris</i>	402997	LC
<i>Gyraulus laevis</i>	430518	VU
<i>Hadena compta</i>	448102	NA
<i>Haitia acuta</i>	430438	NE
<i>Hamearis lucina</i>	440909	VU
<i>Hemaris fuciformis</i>	443647	NA
<i>Hemaris tityus</i>	443640	NA
<i>Hemianax ephippiger</i>	214326	NE
<i>Hesperia comma</i>	440721	NA
<i>Hippeutis complanatus</i>	430484	LC
<i>Hippolais icterina</i>	97334	VU
<i>Hyla arborea</i>	177887	EN
<i>Hyles euphorbiae</i>	443766	NA
<i>Hyles gallii</i>	443793	NA
<i>Iphiclides podalirius</i>	440667	EN
<i>Ischnura elegans</i>	214281	LC
<i>Ischnura pumilio</i>	214286	LC
<i>Issoria lathonia</i>	441740	NA
<i>Ixobrychus minutus</i>	96634	EN
<i>Jordanita globulariae</i>	440643	NA
<i>Jordanita notata</i>	440630	NA
<i>Jynx torquilla</i>	97090	NT

<i>Lacerta agilis</i>	214645	VU
<i>Lanius collurio</i>	97116	LC
<i>Lanius senator</i>	97121	CR
<i>Larus ridibundus</i>	96929	EN
<i>Lasiocampa trifolii</i>	443542	NA
<i>Lasiommata megera</i>	441226	NA
<i>Leptidea sinapis</i>	440900	NA
<i>Leptophyes punctatissima</i>	403278	LC
<i>Lepus europaeus</i>	305525	VU
<i>Lestes viridis</i>	214252	LC
<i>Leucorrhinia caudalis</i>	214380	CR
<i>Leucorrhinia pectoralis</i>	214382	CR
<i>Libelloides coccajus</i>	85237	VU
<i>Libellula depressa</i>	214384	LC
<i>Libellula fulva</i>	214385	LC
<i>Libellula quadrimaculata</i>	214386	LC
<i>Limenitis reducta</i>	441651	EN
<i>Locustella luscinioides</i>	97309	NT
<i>Locustella naevia</i>	97307	NT
<i>Lopinga achine</i>	441223	EN
<i>Lucanus cervus</i>	123296	NA
<i>Luscinia megarhynchos</i>	97174	NT
<i>Lycaena dispar</i>	441190	NA
<i>Lycaena phlaeas</i>	441185	NA
<i>Lycaena tityrus</i>	441198	NA
<i>Lycaena virgaureae</i>	441193	VU
<i>Macroglossum stellatarum</i>	443702	NA
<i>Maniola jurtina</i>	441303	NA
<i>Mecostethus parapleurus</i>	402378	LC
<i>Melanargia galathea</i>	441445	NA
<i>Melanchra persicariae</i>	448252	NA
<i>Melitaea athalia</i>	441611	VU
<i>Melitaea diamina</i>	441597	VU
<i>Melitaea didyma</i>	441594	VU
<i>Melitaea parthenoides</i>	441604	EN
<i>Melitta nigricans</i>	232909	EN
<i>Mergus merganser</i>	96551	VU
<i>Metrioptera bicolor</i>	403499	VU
<i>Metrioptera brachyptera</i>	403500	NT

<i>Microlynyphia impigra</i>	350719	NA
<i>Miliaria calandra</i>	97541	VU
<i>Milvus migrans</i>	96688	LC
<i>Milvus milvus</i>	96689	LC
<i>Minucia lunaris</i>	446631	NA
<i>Motacilla alba</i>	97427	LC
<i>Motacilla cinerea</i>	97426	LC
<i>Muscardinus avellanarius</i>	305717	VU
<i>Muscicapa striata</i>	97159	LC
<i>Musculium lacustre</i>	337909	LC
<i>Mustela erminea</i>	305315	LC
<i>Mustela nivalis</i>	305324	VU
<i>Myotis bechsteinii</i>	305408	NT
<i>Myotis daubentonii</i>	305417	VU
<i>Myotis myotis</i>	305419	EN
<i>Myotis nattereri</i>	305421	NT
<i>Natrix natrix</i>	214751	EN
<i>Noctua fimbriata</i>	448845	NA
<i>Noctua pronuba</i>	448843	NA
<i>Notonecta glauca</i>	450468	NA
<i>Nyctalus noctula</i>	305429	VU
<i>Nymphalis antiopa</i>	441666	VU
<i>Nymphalis polychloros</i>	441667	VU
<i>Odezia atrata</i>	444521	NA
<i>Oecanthus pellucens</i>	402913	LC
<i>Oedipoda caerulea</i>	402365	NT
<i>Omocestus rufipes</i>	402488	NT
<i>Omocestus viridulus</i>	402492	LC
<i>Onychogomphus forcipatus</i>	214340	EN
<i>Opisthograptis luteolata</i>	446005	NA
<i>Orgyia antiqua</i>	447075	NA
<i>Oriolus oriolus</i>	97124	LC
<i>Orthetrum albistylum</i>	214388	NA
<i>Orthetrum brunneum</i>	214389	LC
<i>Orthetrum cancellatum</i>	214390	LC
<i>Orthetrum coerulescens</i>	214392	NT
<i>Papilio machaon</i>	440671	NA
<i>Pararge aegeria</i>	441236	NA
<i>Parnassius apollo</i>	440702	VU
<i>Parus ater</i>	97272	LC
<i>Parus caeruleus</i>	97274	LC

<i>Parus major</i>	97276	LC
<i>Parus palustris</i>	97267	LC
<i>Passer domesticus</i>	97437	LC
<i>Passer montanus</i>	97441	LC
<i>Phalera bucephala</i>	446378	NA
<i>Phaneroptera falcata</i>	403293	VU
<i>Phengaris arion</i>	441074	VU
<i>Phoenicurus ochruros</i>	97184	LC
<i>Phoenicurus phoenicurus</i>	97185	NT
<i>Phragmatobia fuliginosa</i>	447012	NA
<i>Phragmatobia luctifera</i>	447014	NA
<i>Phylloscopus bonelli</i>	97367	LC
<i>Phylloscopus collybita</i>	97373	LC
<i>Phylloscopus trachilus</i>	97375	VU
<i>Pica pica</i>	97133	LC
<i>Picus canus</i>	97092	VU
<i>Picus viridis</i>	97093	LC
<i>Pieris brassicae</i>	440880	NA
<i>Pieris napi</i>	440887	NA
<i>Pieris rapae</i>	440885	NA
<i>Plagodis dolabraria</i>	446227	NA
<i>Planorbis carinatus</i>	430469	LC
<i>Platycleis albopunctata</i>	403578	NT
<i>Plebejus argus</i>	441032	VU
<i>Plebejus idas</i>	441036	VU
<i>Plecotus auritus</i>	305438	VU
<i>Podarcis muralis</i>	214670	LC
<i>Podiceps cristatus</i>	96561	LC
<i>Polygonia c-album</i>	441674	NA
<i>Polyommatus bellargus</i>	440948	NA
<i>Polyommatus coridon</i>	440950	VU
<i>Polyommatus icarus</i>	440934	NA
<i>Polyommatus thersites</i>	440933	VU
<i>Pontia daplidice</i>	440877	EN
<i>Prunella modularis</i>	97429	LC
<i>Pseudochorthippus montanus</i>	402589	VU
<i>Psophus stridulus</i>	402353	VU
<i>Pteronemobius heydenii</i>	402927	VU
<i>Pupilla muscorum</i>	430797	NA
<i>Pyrgus malvae</i>	440751	VU

<i>Pyrrhosoma nymphula</i>	214292	LC
<i>Pyrrhula pyrrhula</i>	97495	LC
<i>Rallus aquaticus</i>	96767	LC
<i>Rana esculenta</i>	177908	NT
<i>Rana lessonae</i>	177916	NT
<i>Rana ridibunda</i>	177919	NE
<i>Rana temporaria</i>	177921	LC
<i>Regulus ignicapillus</i>	97297	LC
<i>Regulus regulus</i>	97294	LC
<i>Rhizotrogus aestivus</i>	257568	NA
<i>Rhodostrophia vibicaria</i>	444265	NA
<i>Riparia riparia</i>	97282	VU
<i>Ruspolia nitidula</i>	403186	NT
<i>Salamandra salamandra</i>	177846	VU
<i>Satyrium pruni</i>	441169	CR
<i>Saxicola rubetra</i>	97189	VU
<i>Saxicola rubicola</i>	97192	NT
<i>Scolopax rusticola</i>	96827	VU
<i>Segmentina nitida</i>	430462	VU
<i>Serinus serinus</i>	97462	LC
<i>Siona lineata</i>	446008	NA
<i>Sitta europaea</i>	97254	LC
<i>Somatochlora flavomaculata</i>	214366	LC
<i>Somatochlora metallica</i>	214368	LC
<i>Sphingonotus caeruleus</i>	402321	VU
<i>Sphinx ligustri</i>	443872	NA
<i>Spialia sertorius</i>	440779	NA
<i>Spilosoma lubricipeda</i>	446997	NA
<i>Spilosoma lutea</i>	446996	NA
<i>Stenobothrus lineatus</i>	402447	LC
<i>Sterna hirundo</i>	96959	NT
<i>Stethophyma grossum</i>	402319	VU
<i>Streptopelia turtur</i>	97013	NT
<i>Strix aluco</i>	97047	LC
<i>Sturnus vulgaris</i>	97244	LC
<i>Sylvia atricapilla</i>	97352	LC
<i>Sylvia borin</i>	97351	NT
<i>Sylvia communis</i>	97350	NT
<i>Symplectma fusca</i>	214254	LC
<i>Sympetrum depressiusculum</i>	214400	VU
<i>Sympetrum flaveolum</i>	214401	EN

<i>Sympetrum fonscolombii</i>	214402	NE
<i>Sympetrum pedemontanum</i>	214406	CR
<i>Sympetrum sanguineum</i>	214407	LC
<i>Sympetrum striolatum</i>	214409	LC
<i>Sympetrum vulgatum</i>	214410	LC
<i>Tachybaptus ruficollis</i>	96559	VU
<i>Tetragnatha extensa</i>	353100	NA
<i>Tetralonia malvae</i>	233474	NA
<i>Tetrix bipunctata</i>	402277	NT
<i>Thecla betulae</i>	441157	NA

<i>Thera juniperata</i>	444646	NA
<i>Thymelicus sylvestris</i>	440726	NA
<i>Trachys troglodytiformis</i>	102594	NA
<i>Trichodes alvearius</i>	187146	NA
<i>Tringa ochropus</i>	96850	LC
<i>Triphosa dubitata</i>	445159	NA
<i>Triturus alpestris</i>	177854	LC
<i>Triturus cristatus</i>	177858	EN
<i>Triturus helveticus</i>	177861	VU
<i>Triturus vulgaris</i>	177866	EN
<i>Troglodytes troglodytes</i>	97263	LC
<i>Truncatellina cylindrica</i>	431271	VU

<i>Turdus merula</i>	97226	LC
<i>Turdus philomelos</i>	97234	LC
<i>Turdus pilaris</i>	97228	VU
<i>Tyto alba</i>	97035	NT
<i>Vallonia excentrica</i>	431110	NA
<i>Vanellus vanellus</i>	96893	CR
<i>Vanessa atalanta</i>	441684	NA
<i>Vanessa cardui</i>	441686	NA
<i>Vertigo moulinsiana</i>	431209	EN
<i>Vipera aspis</i>	214763	CR
<i>Vipera berus</i>	214764	EN
<i>Vulpes vulpes</i>	305294	LC
<i>Xerolenta obvia</i>	428799	NT

<i>Xestia c-nigrum</i>	448764	NA
<i>Xestia triangulum</i>	448766	NA
<i>Xylena exsoleta</i>	447671	NA
<i>Zebrina detrita</i>	425370	VU
<i>Zootoca vivipara</i>	214687	LC
<i>Zygaena carniolica</i>	440326	NA
<i>Zygaena filipendulae</i>	440536	NA
<i>Zygaena loti</i>	440360	NA
<i>Zygaena purpuralis</i>	440279	NA